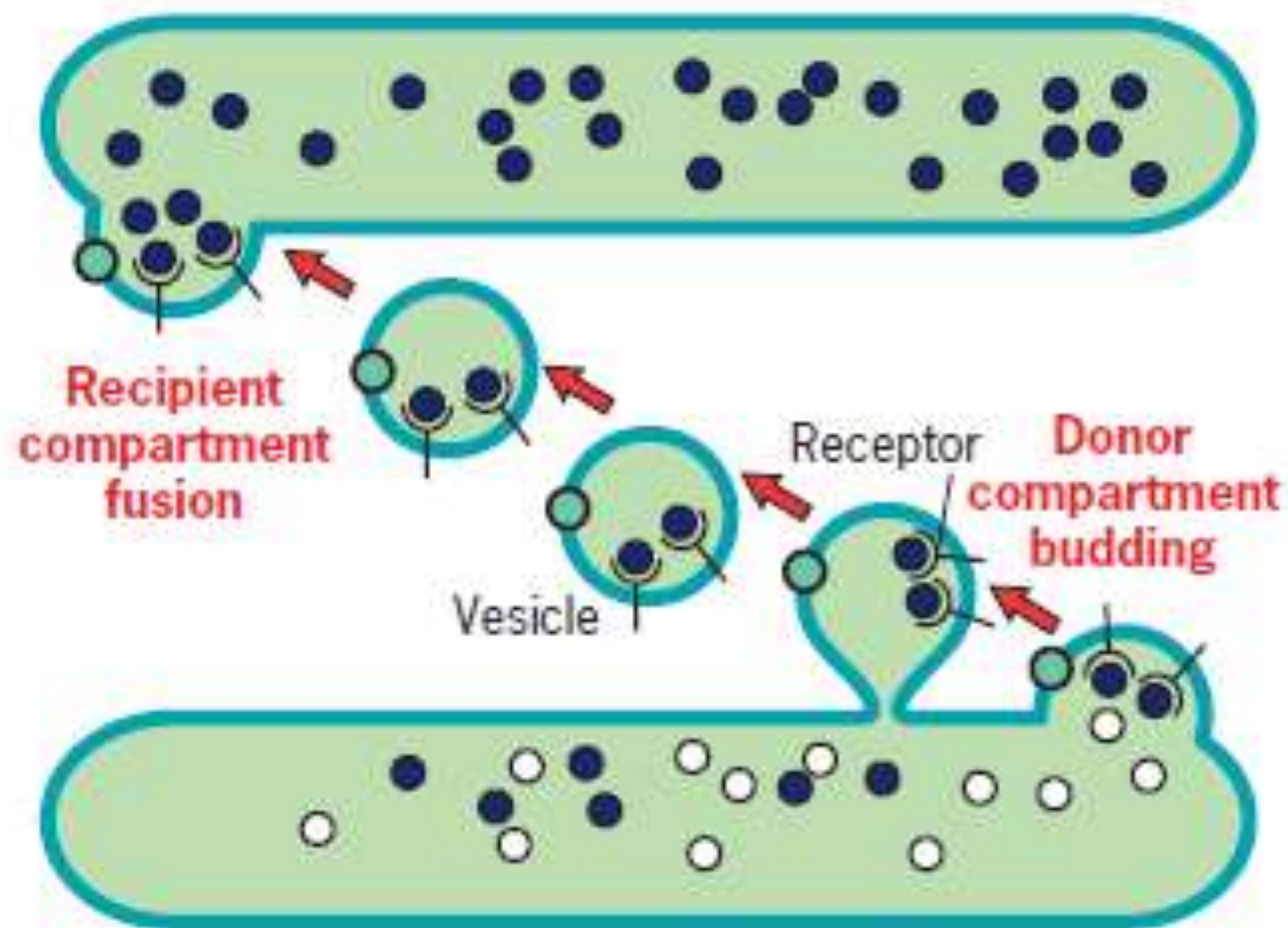


Vesicular trafficking and cellular Transport

Dr. Sudipta Chakraborty
Assistant Professor
PG Department of Microbiology
Bidhannagar College



(a)

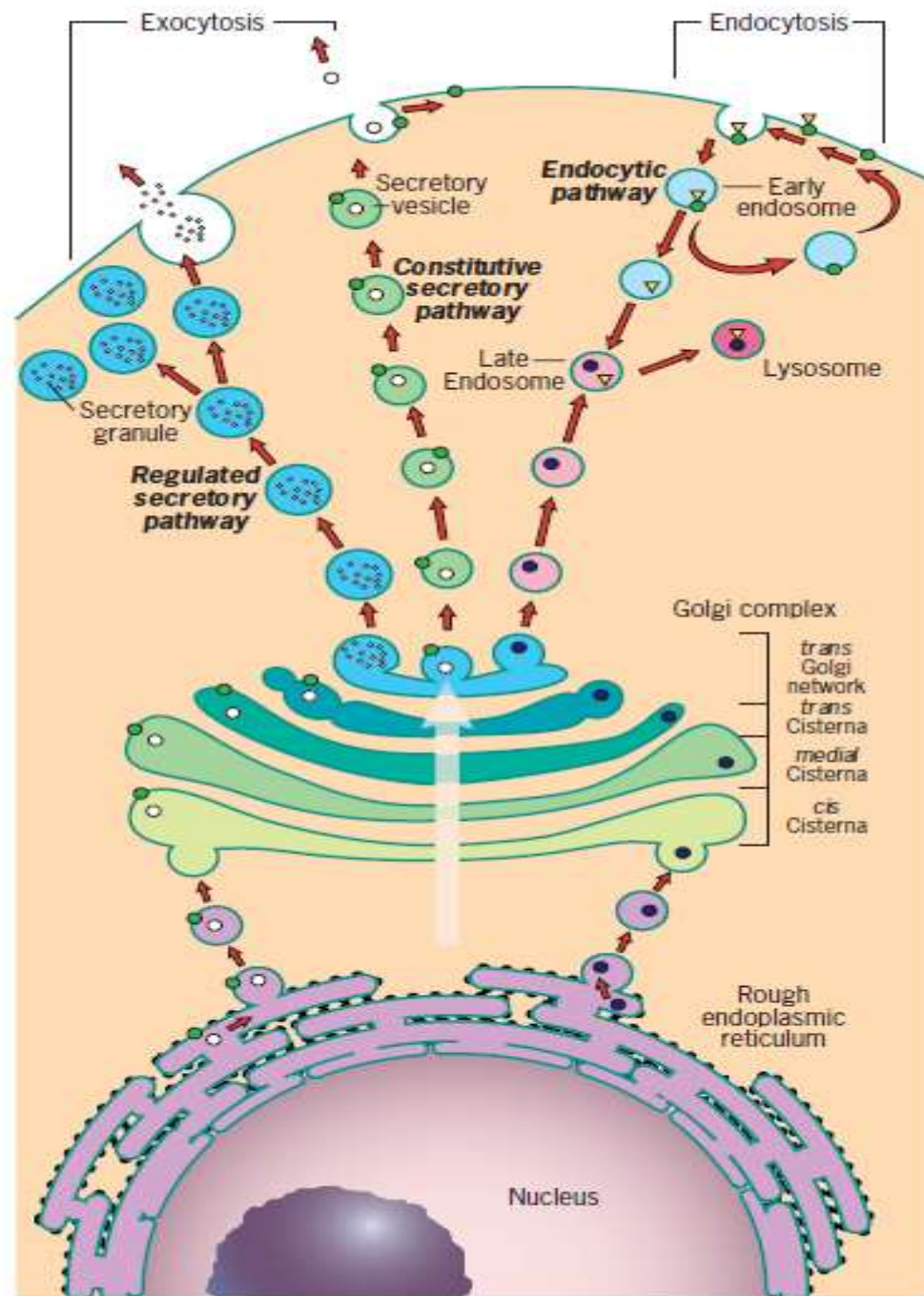
Endomembrane system

Biosynthetic pathway

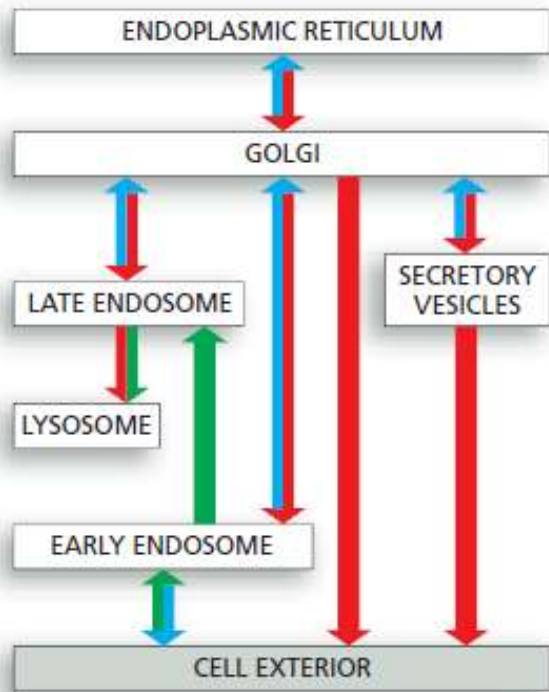
Secretory pathway

Constitutive secretion

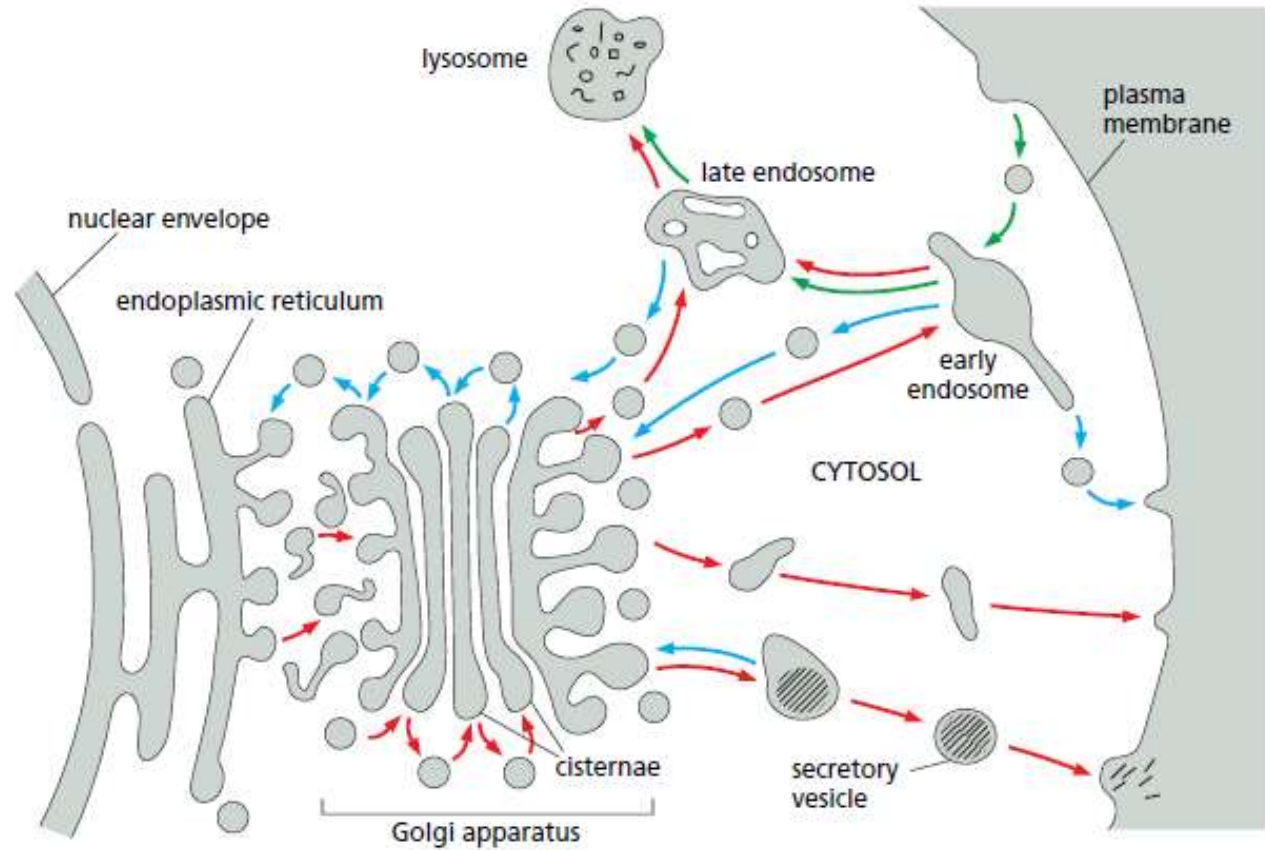
Regulated secretion



(b)

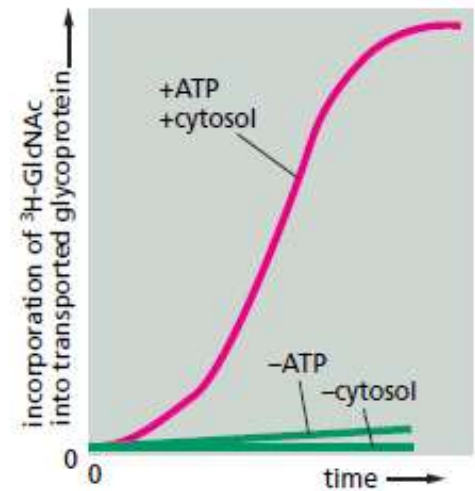
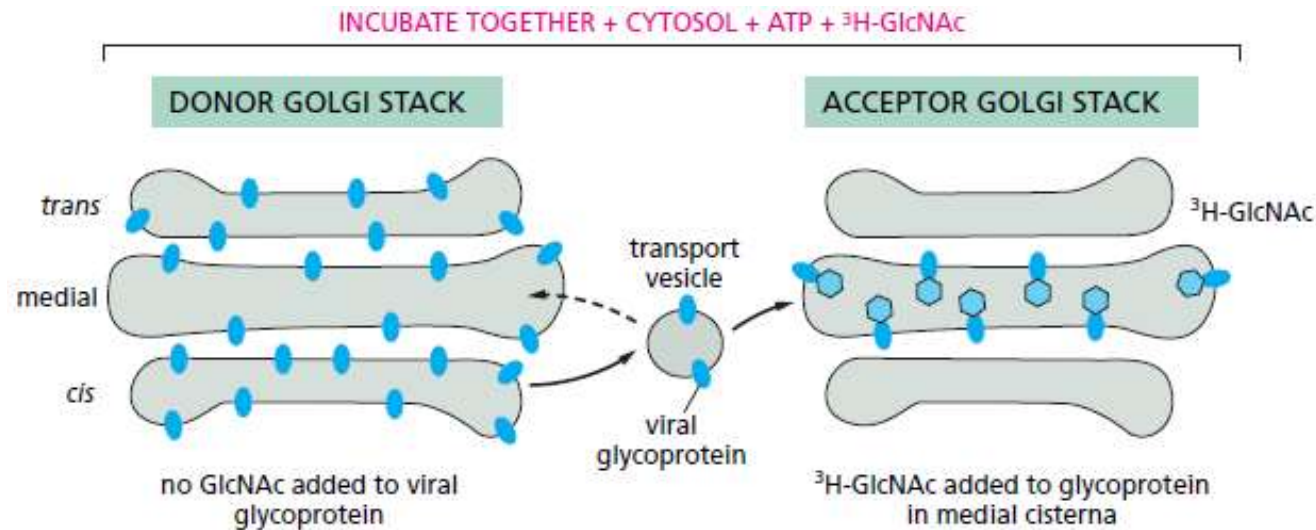


(A)



(B)

CELL-FREE SYSTEMS FOR STUDYING THE COMPONENTS AND MECHANISM OF VESICULAR TRANSPORT



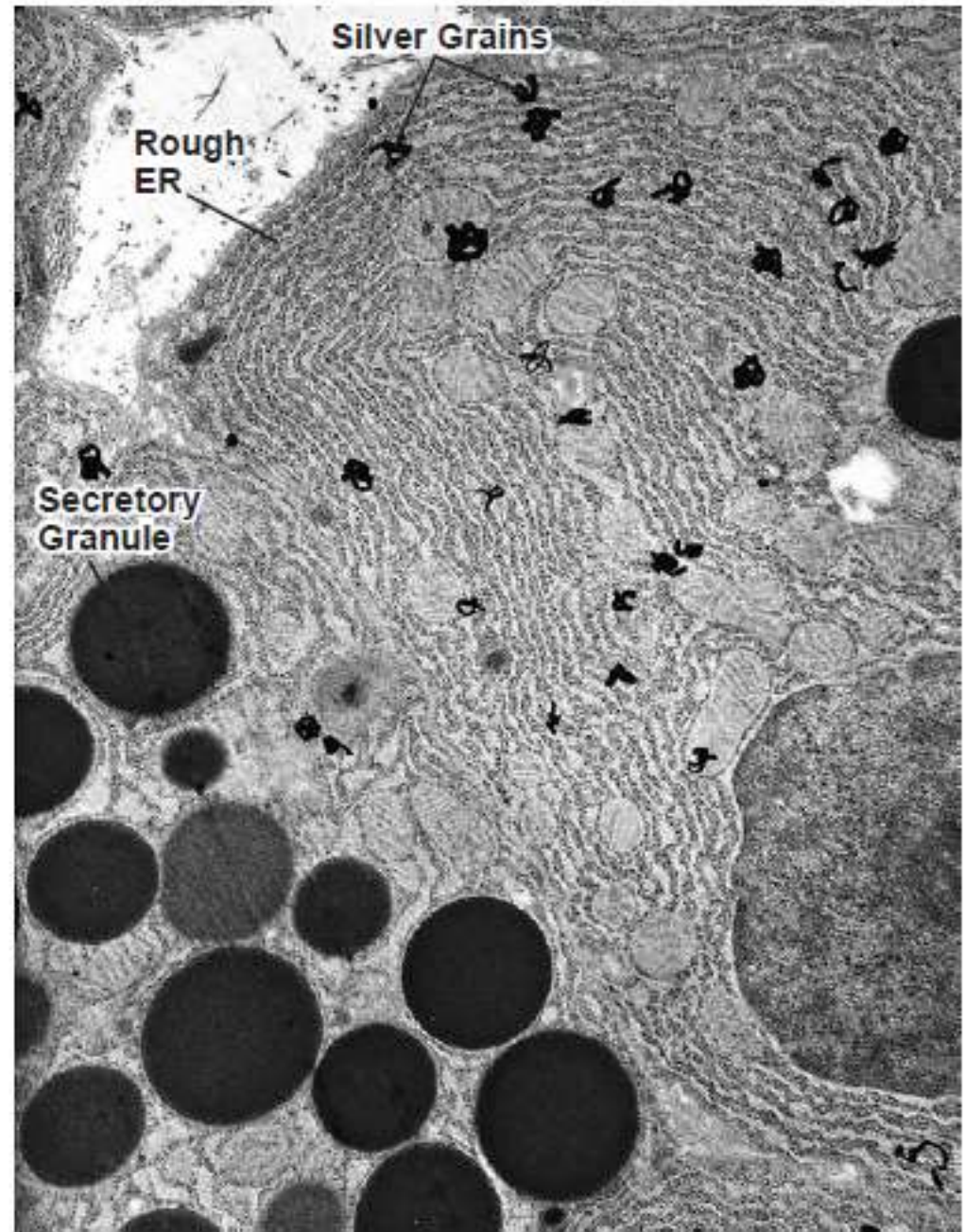
GlcNAc transferase 1 mutant and infected

Wt but uninfected

GENETIC APPROACHES FOR STUDYING VESICULAR TRANSPORT

Multi copy suppression

Autoradiography And Pulse Chase labelling

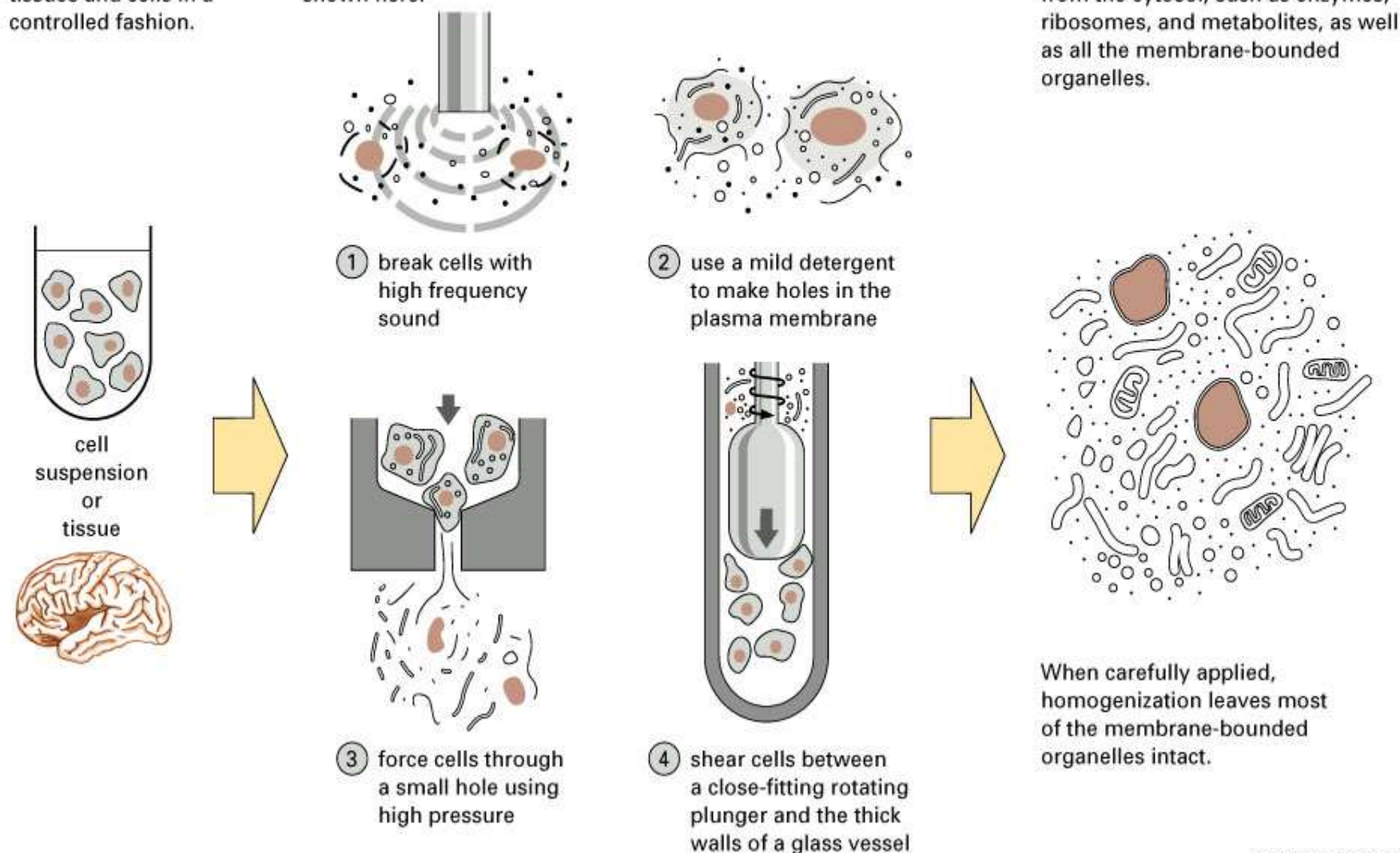


(a)

BREAKING CELLS AND TISSUES

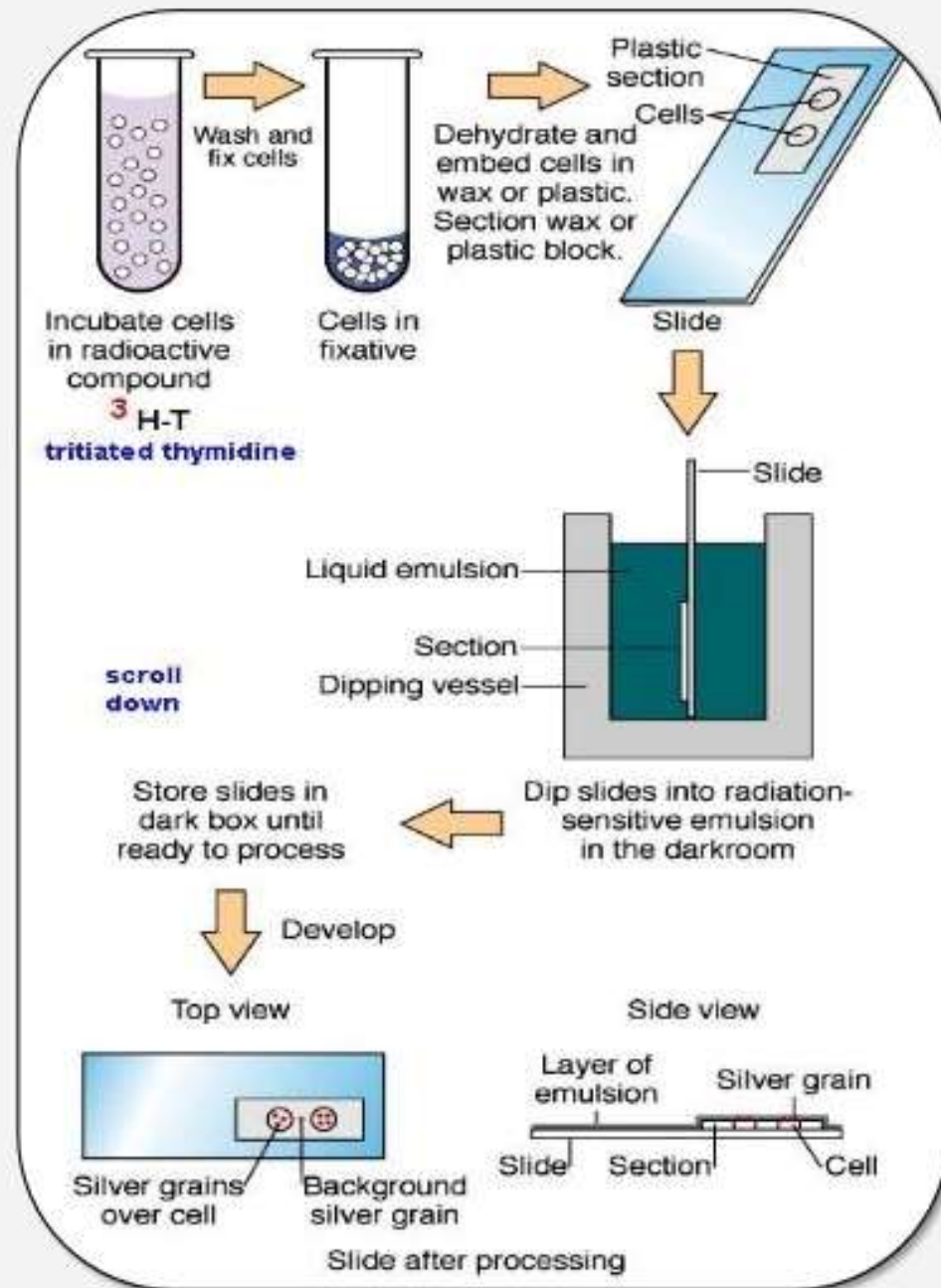
The first step in the purification of most proteins is to disrupt tissues and cells in a controlled fashion.

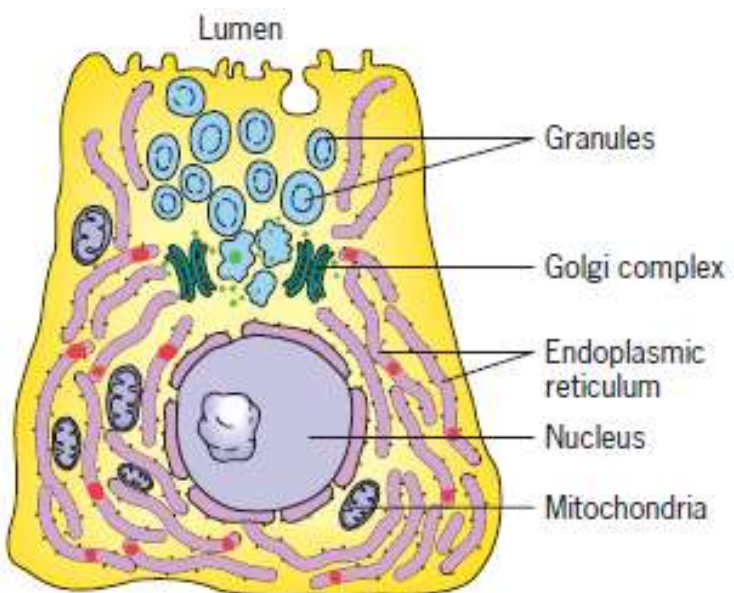
Using gentle mechanical procedures, called homogenization, the plasma membranes of cells can be ruptured so that the cell contents are released. Four commonly used procedures are shown here.



AUTORADIOGRAPHY

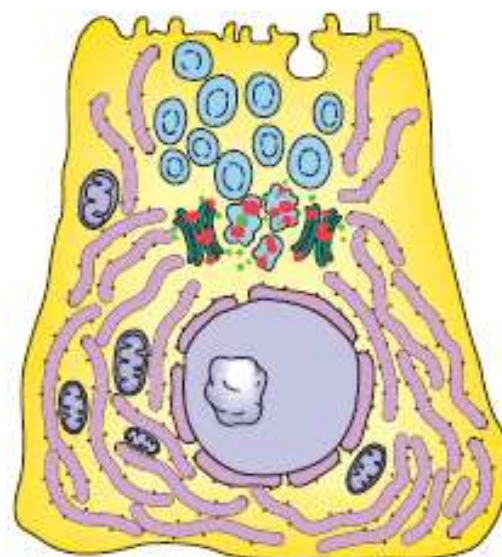
- Radioisotopes are taken up selectively by cells to be studied
- Exposure of photographic film to their emitted radiation reveal presence of such isotopes in the vicinity of these target cells
- Silver bromide crystals in emulsion detect radiation, that reduce them to visible black granules.





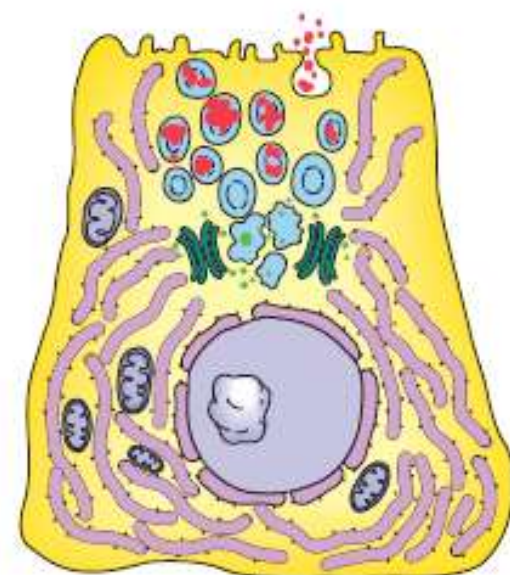
3 min

(b)



20 min

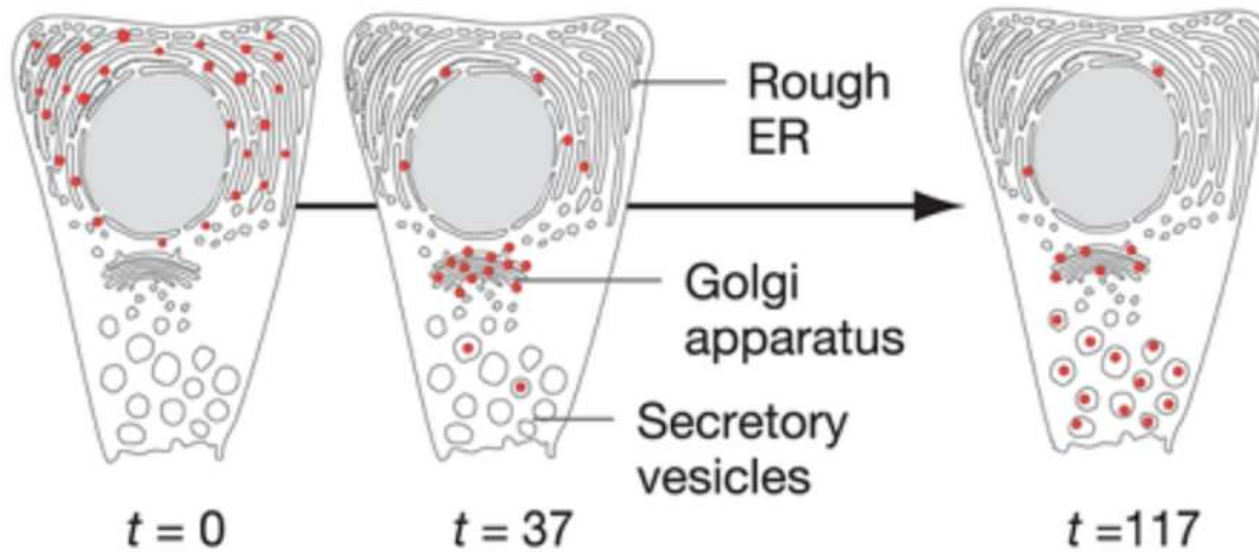
(c)



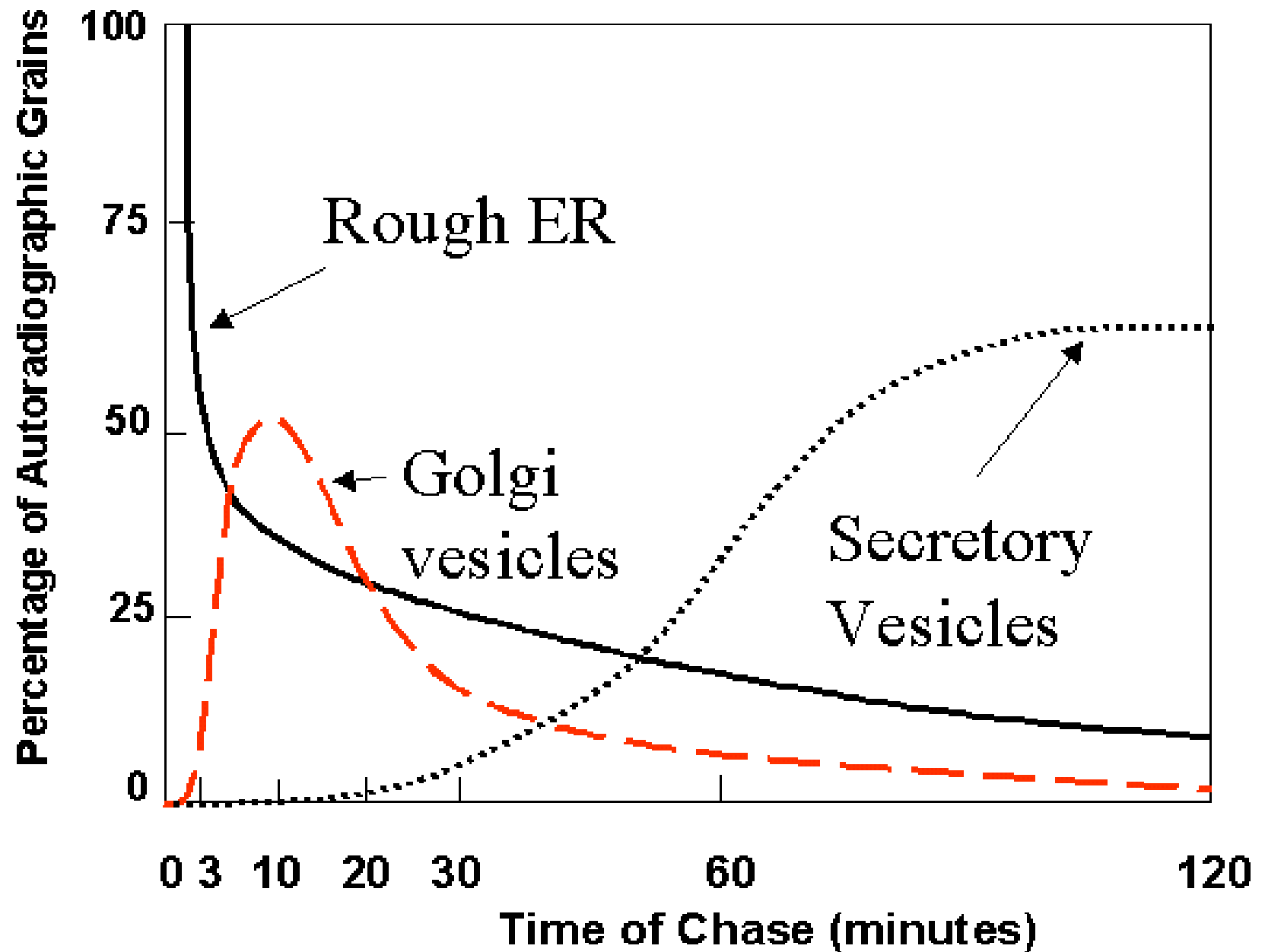
120 min

(d)

Based on the results from this pulse-chase experiment, where are most secreted proteins synthesized?



- ☐ in the rough ER
- ☐ in the Golgi apparatus
- ☐ in the secretory vesicles
- ☐ The answer cannot be determined using this assay.

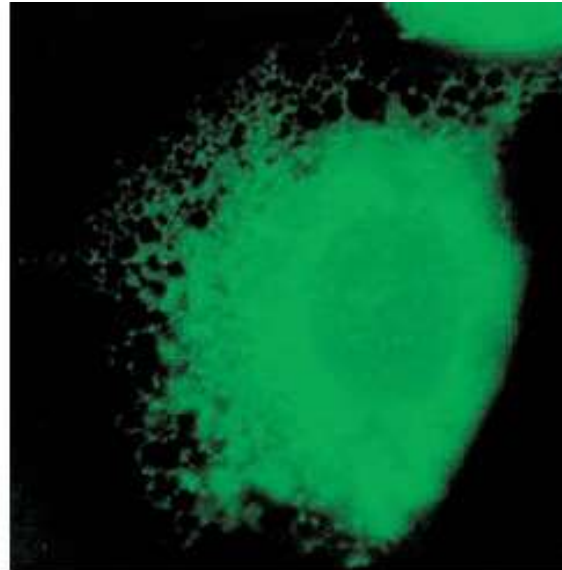


GFP protein

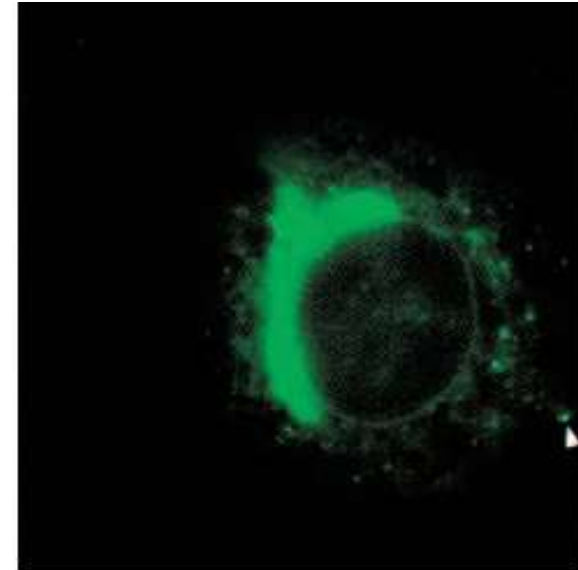
Viral genome contains temperature-sensitive VSVG gene fused to GFP gene



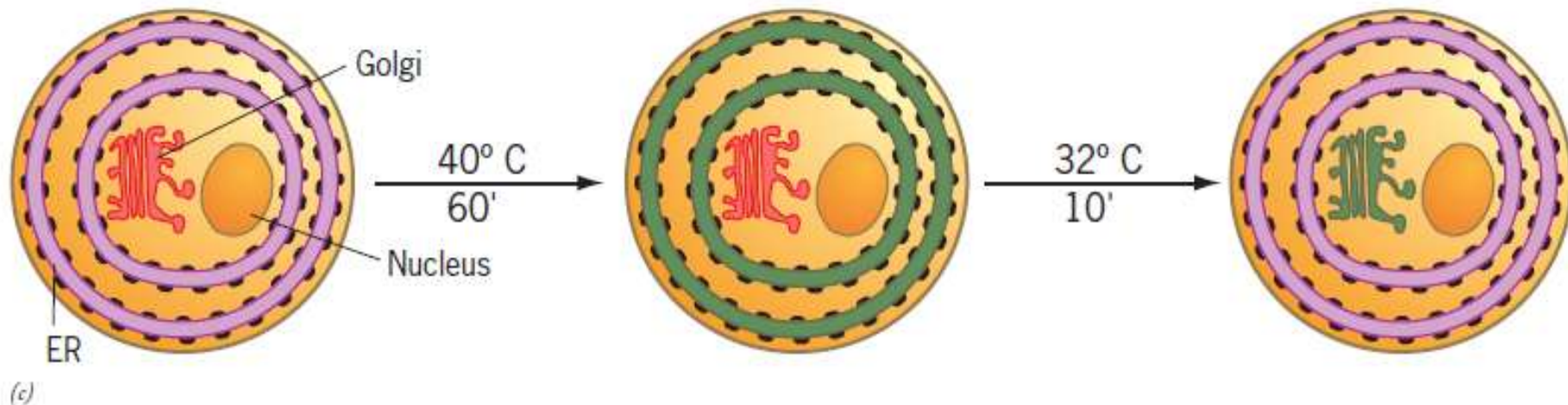
+



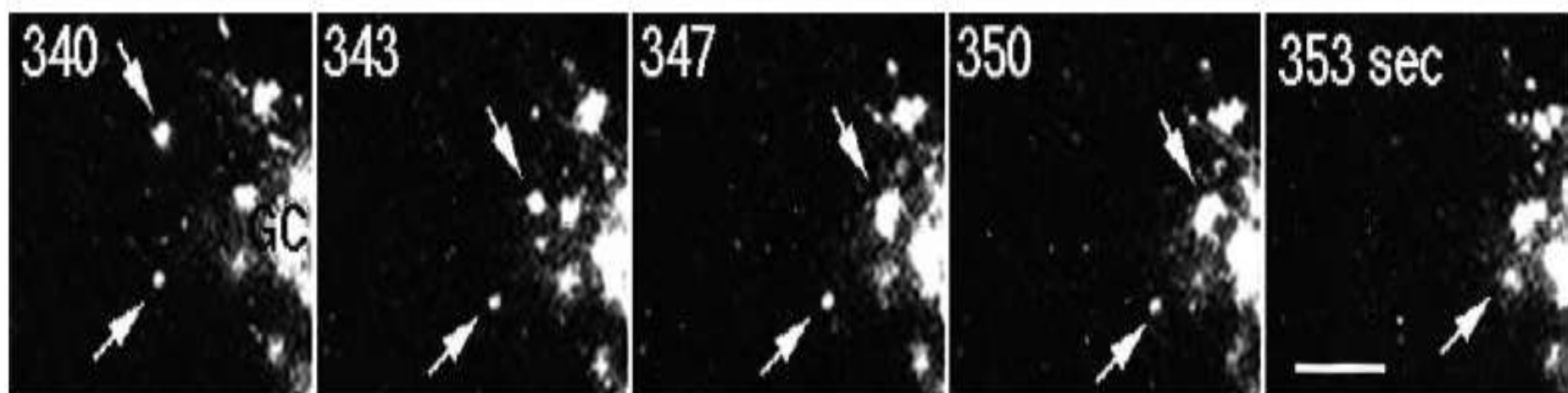
(a)



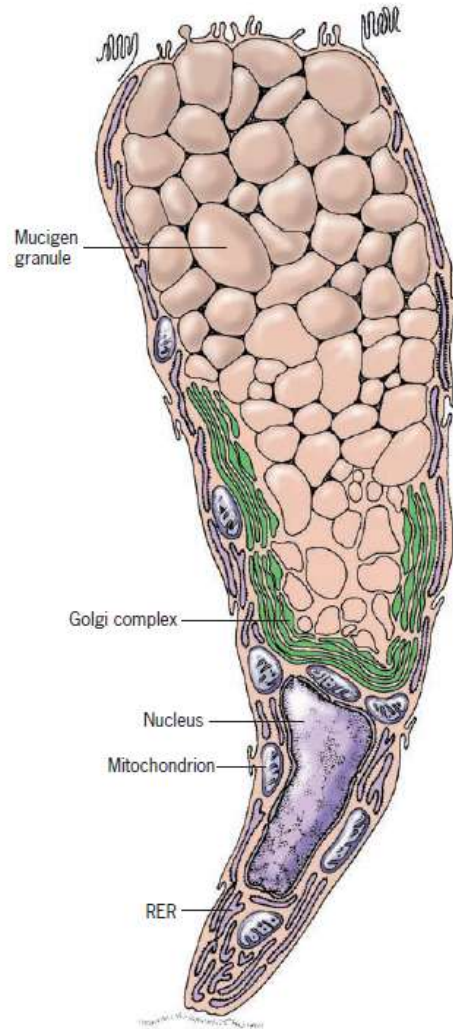
(b)



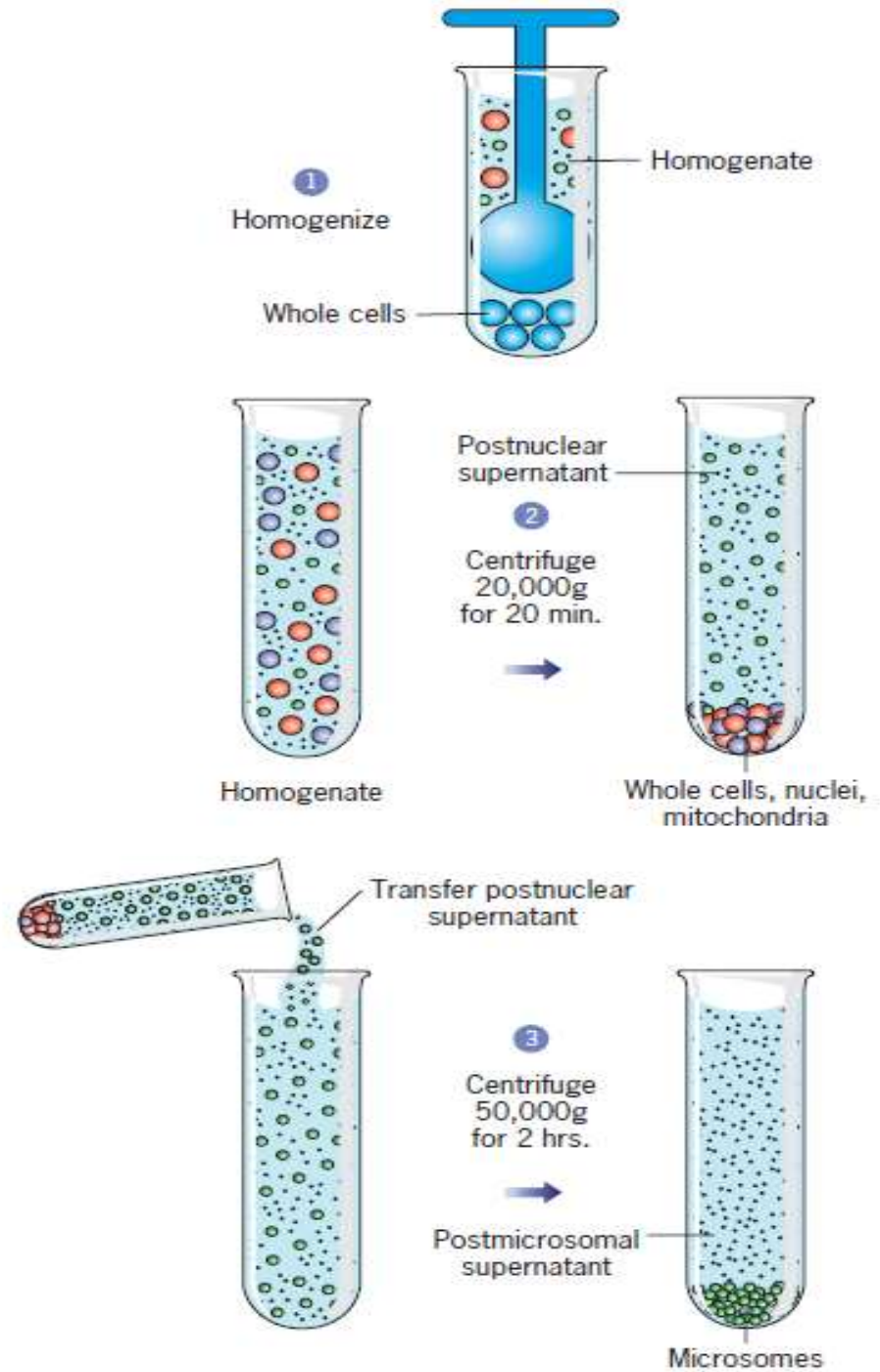
(c)



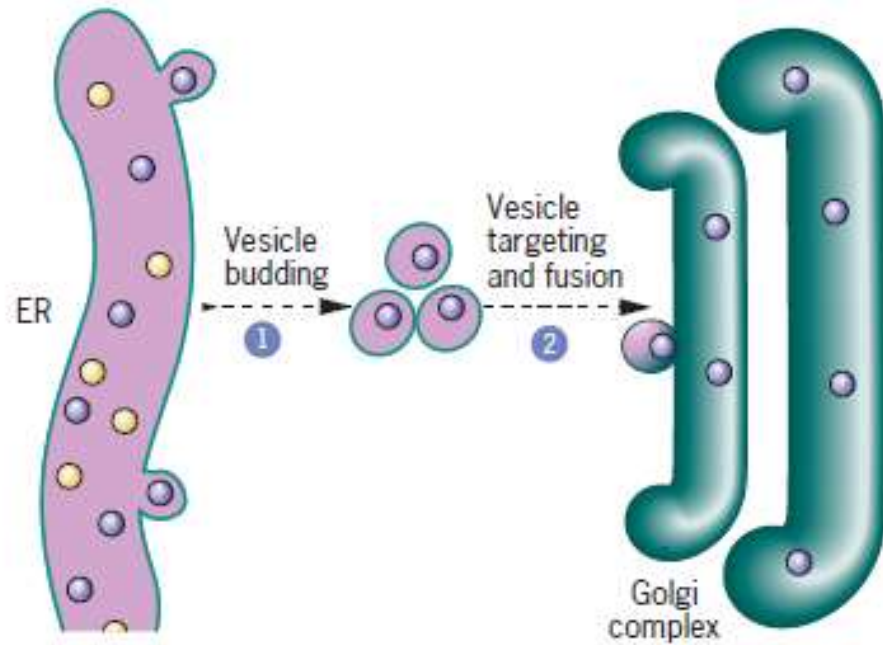
Subcellular Fractionation



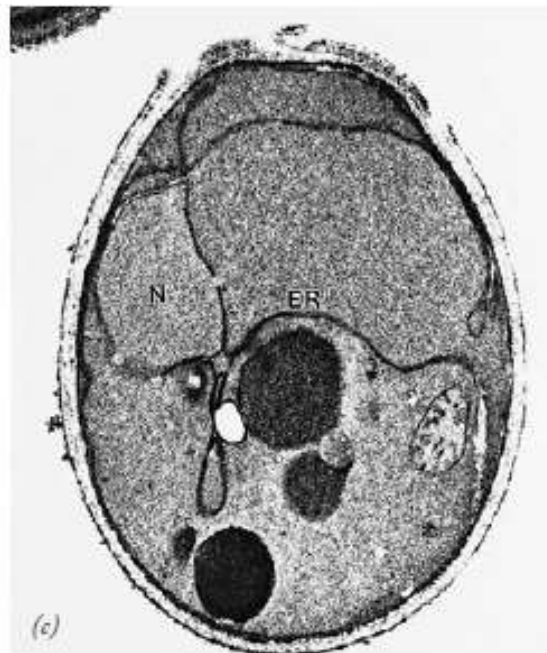
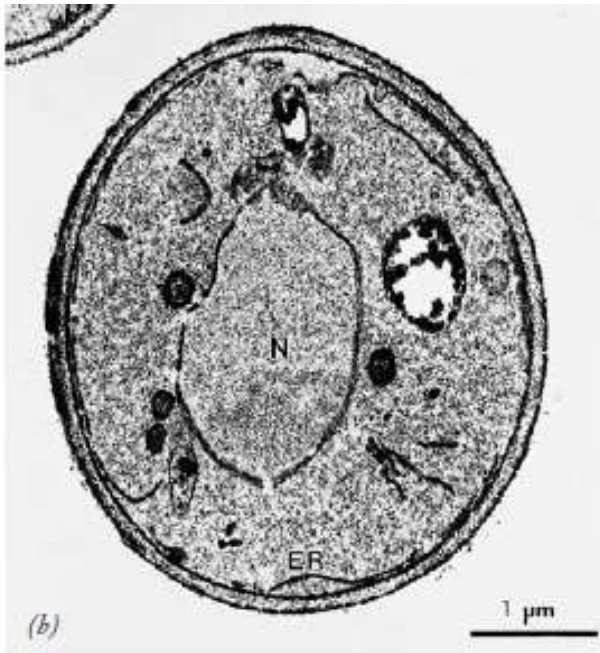
(a)



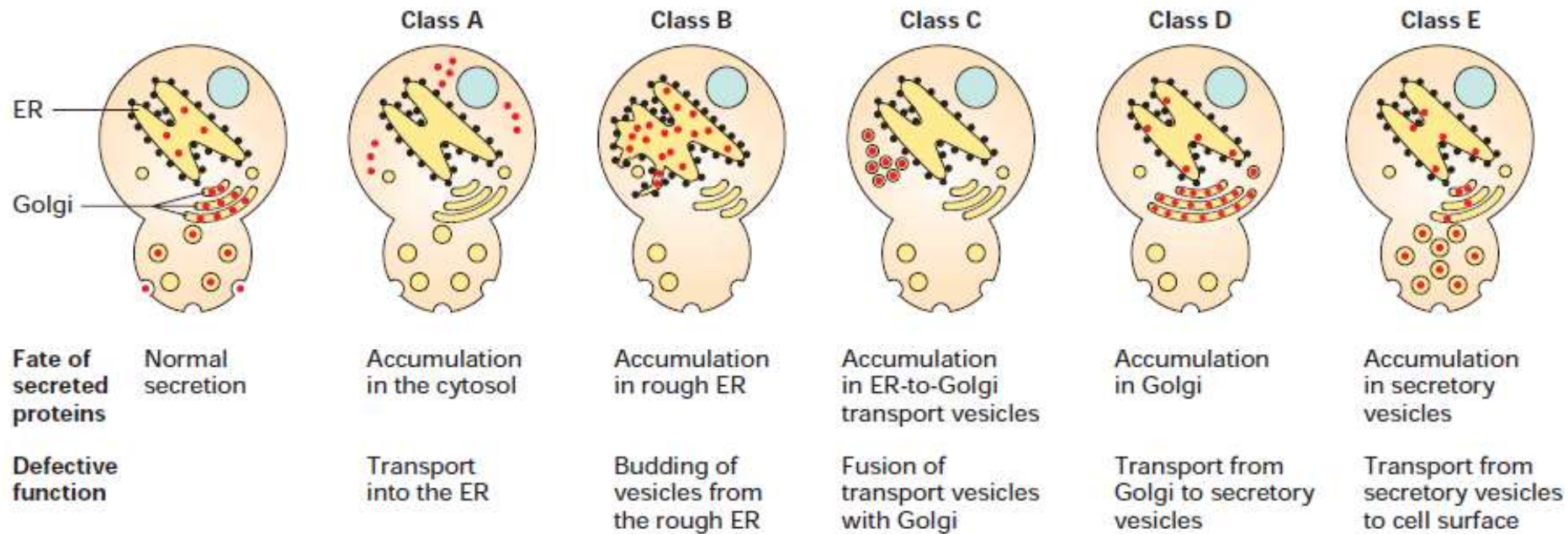
(a)

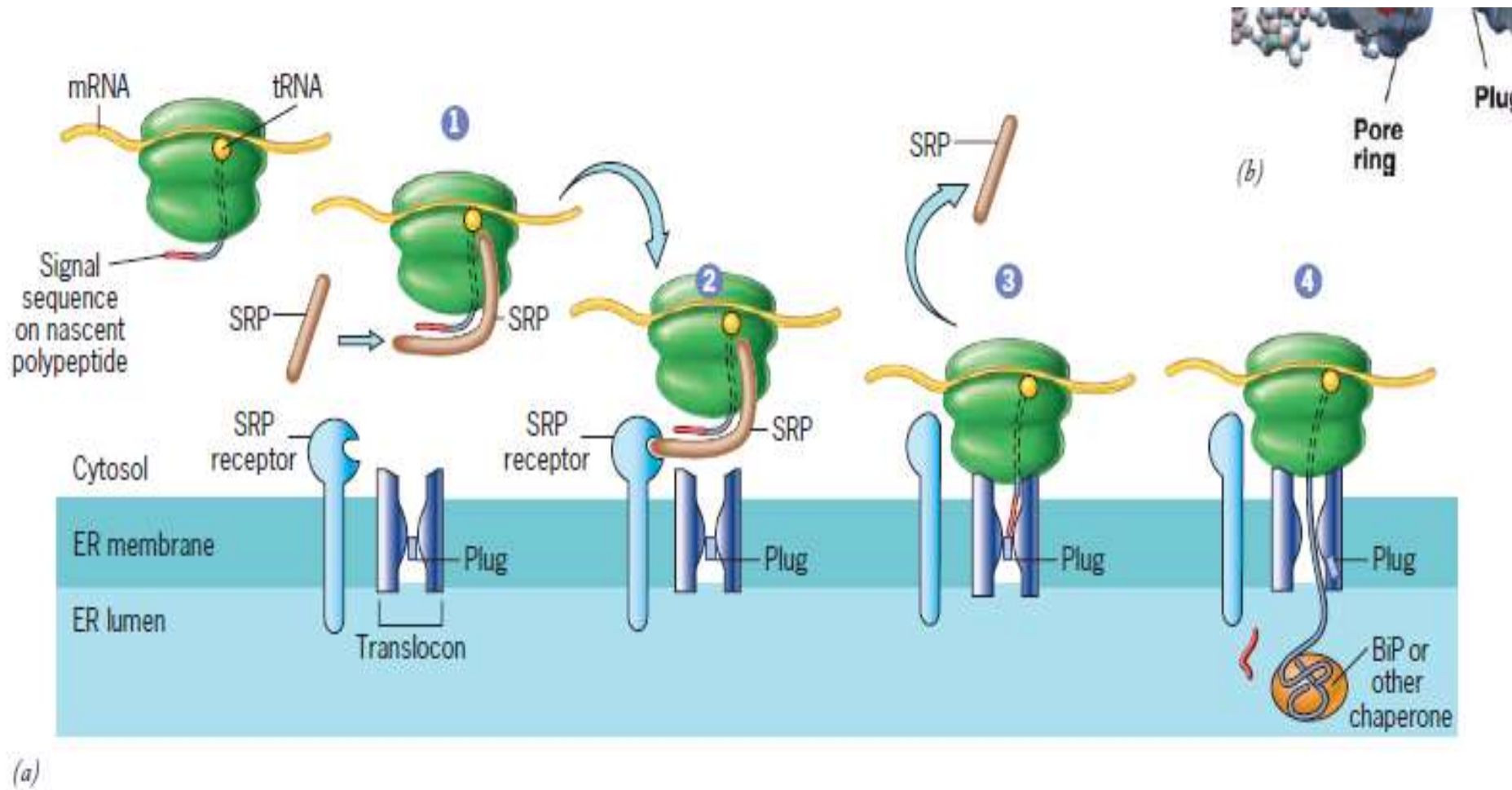


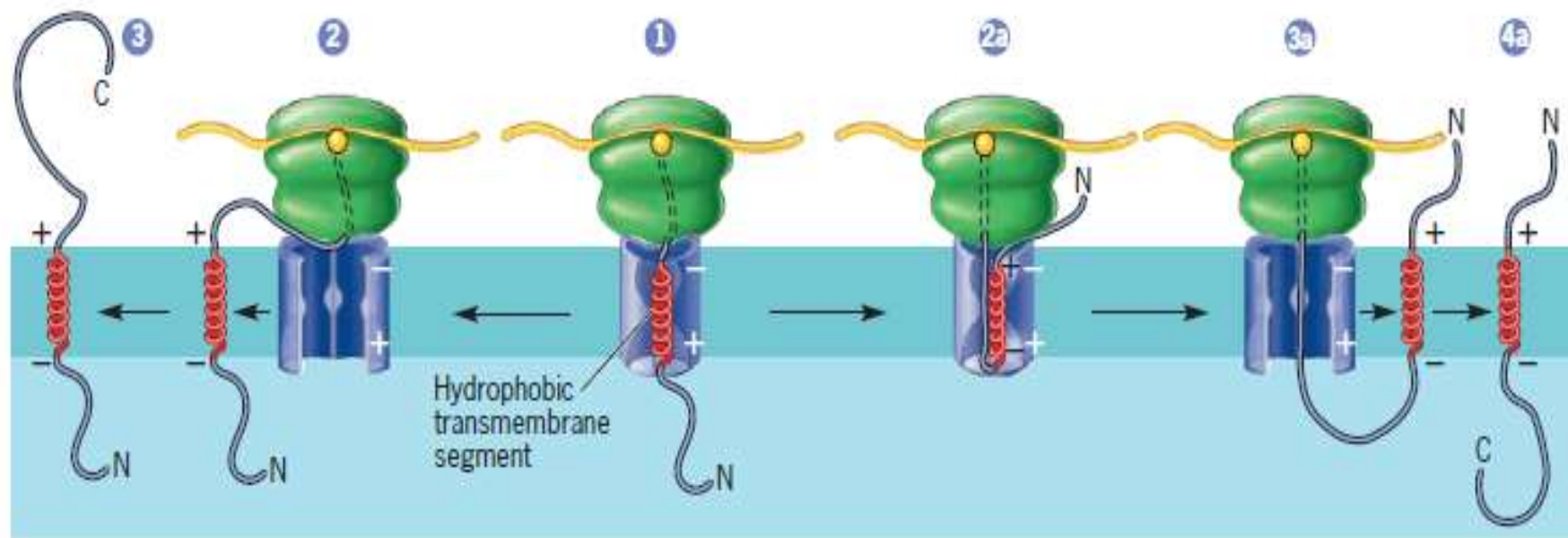
The use of genetic mutants in the study of secretion



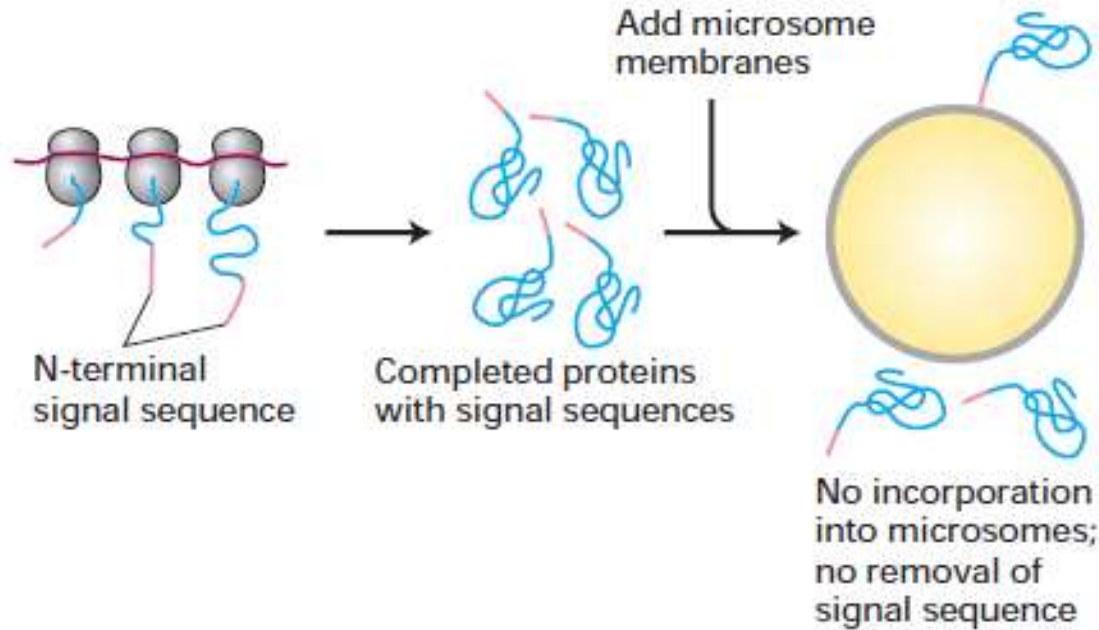
Phenotypes of yeast *sec* mutants identified stages in the secretory pathway



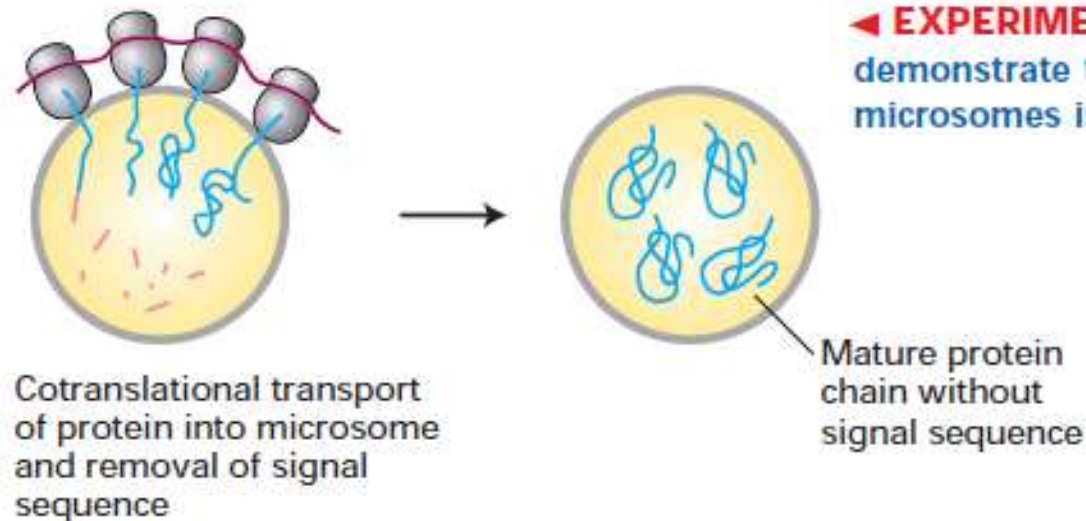




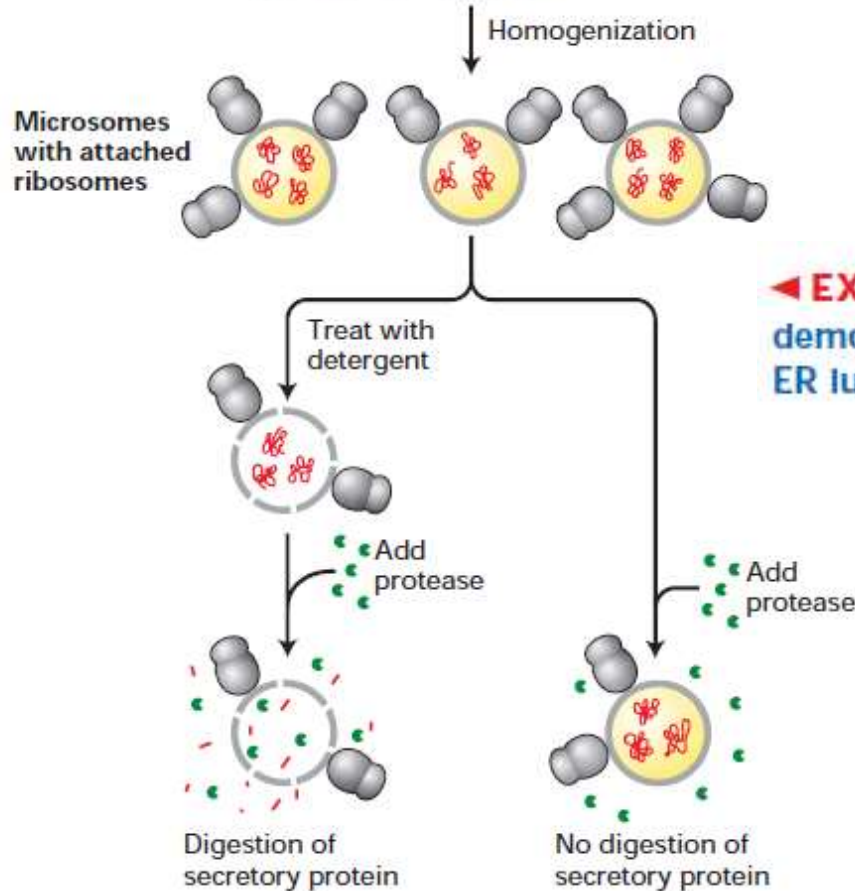
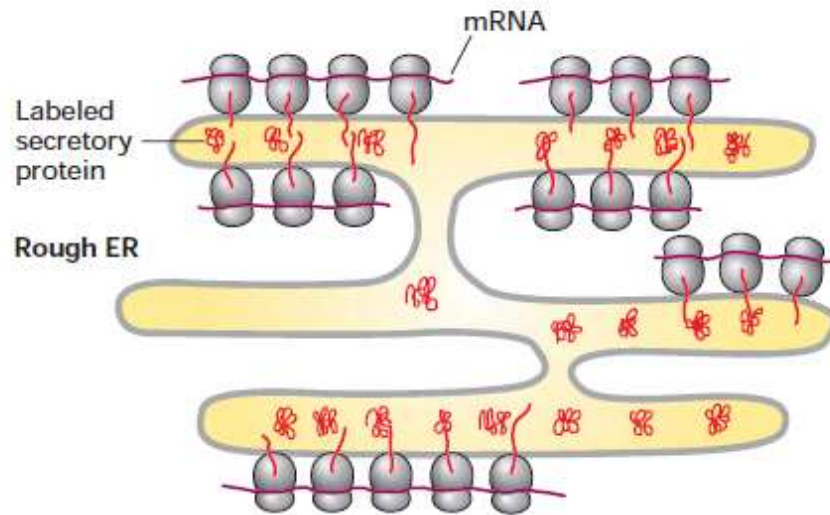
(a) Cell-free protein synthesis; no microsomes present



(b) Cell-free protein synthesis; microsomes present

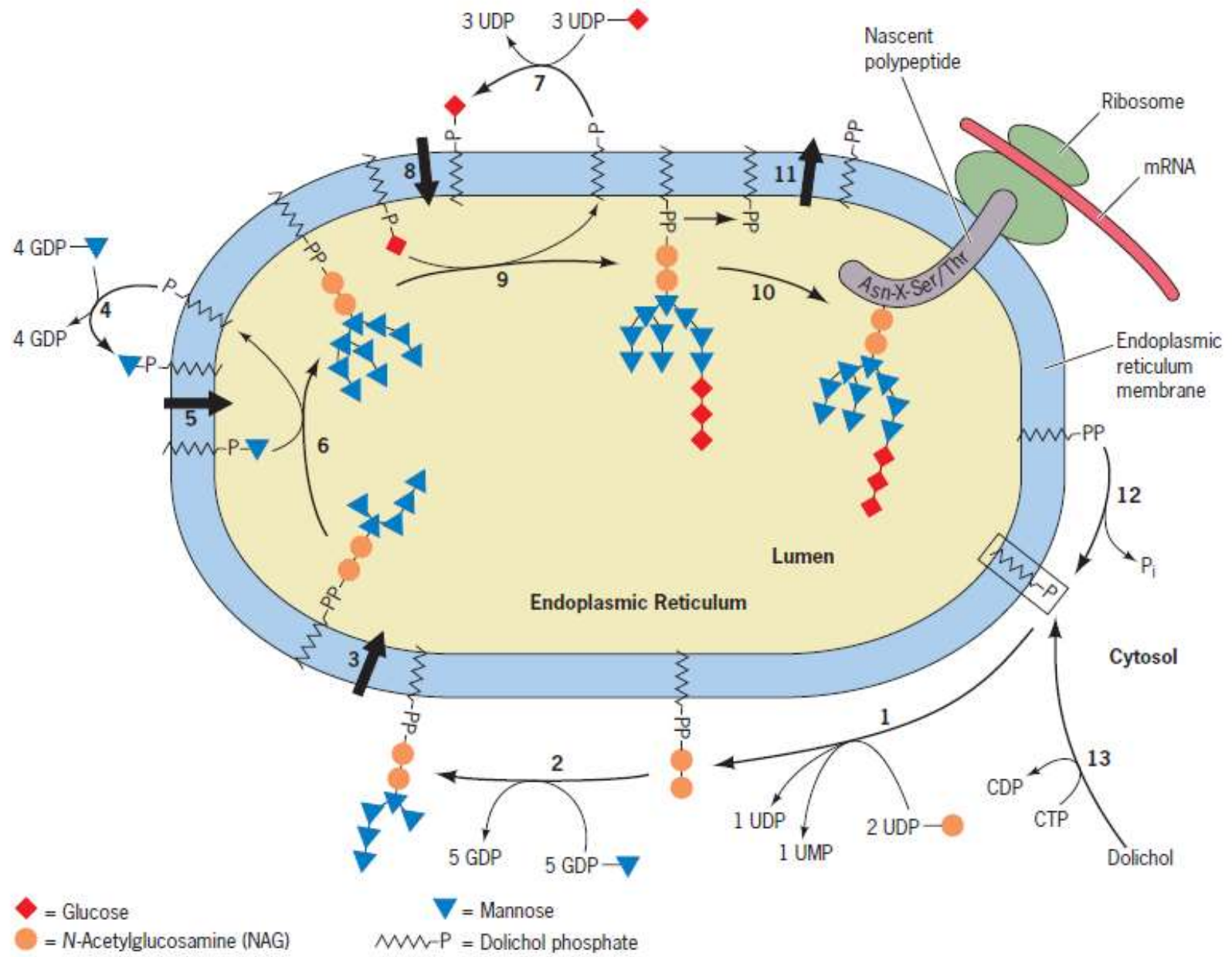


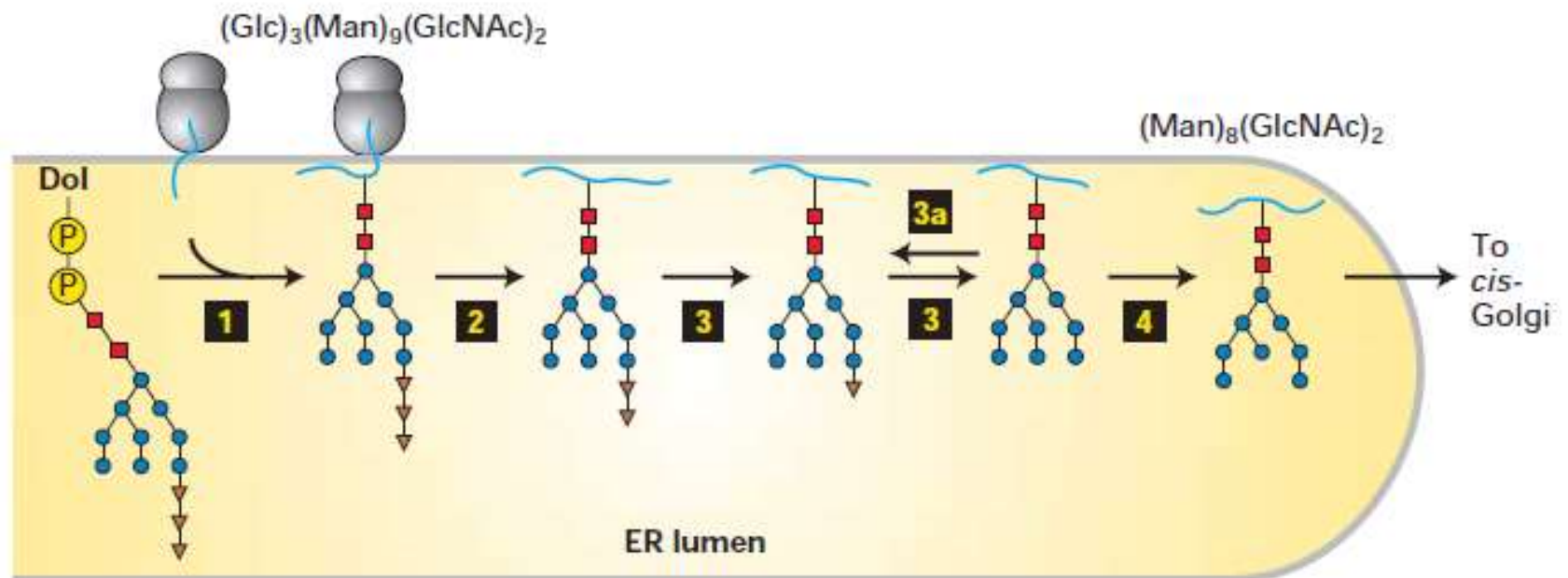
◀ **EXPERIMENTAL FIGURE 16-4** Cell-free experiments demonstrate that translocation of secretory proteins into microsomes is coupled to translation. Treatment of



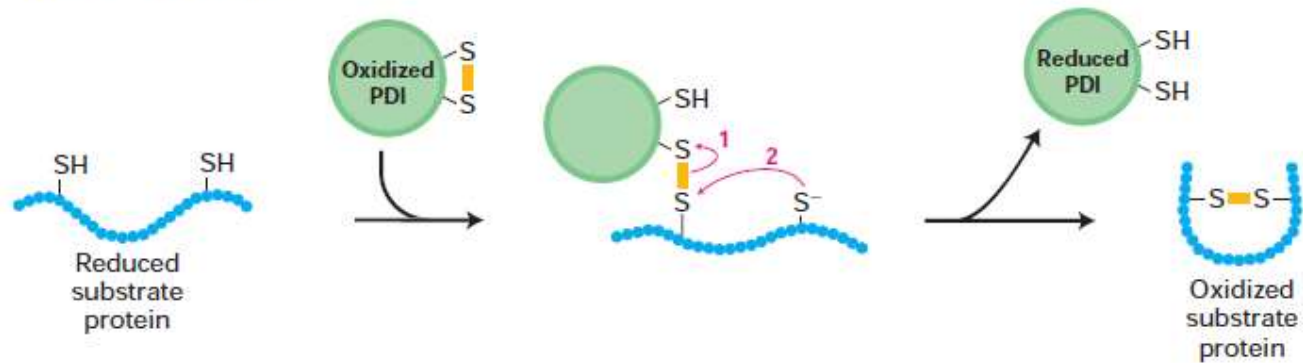
◀ **EXPERIMENTAL FIGURE 16-3** Labeling experiments demonstrate that secretory proteins are localized to the ER lumen shortly after synthesis. Cells are incubated for a

Steps in the synthesis of the core portion of an *N-linked* oligosaccharide in the rough ER

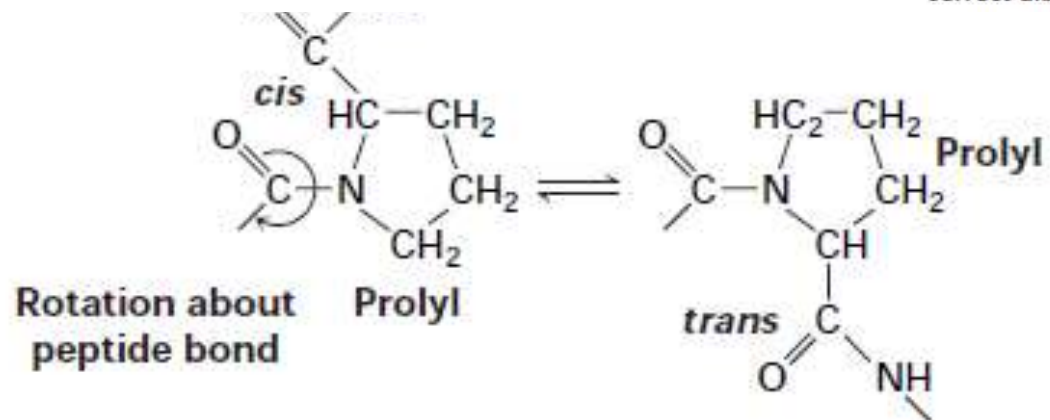
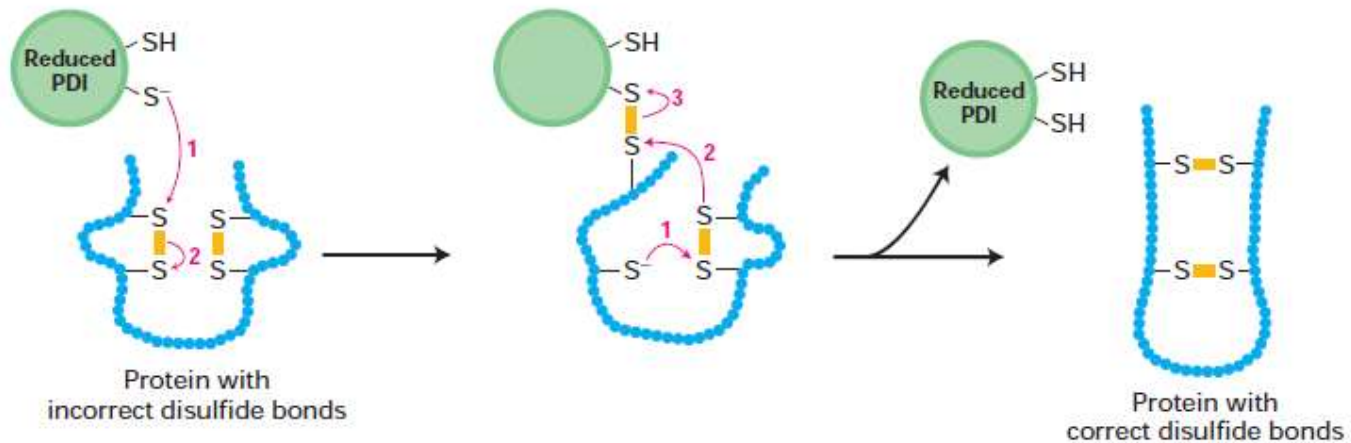


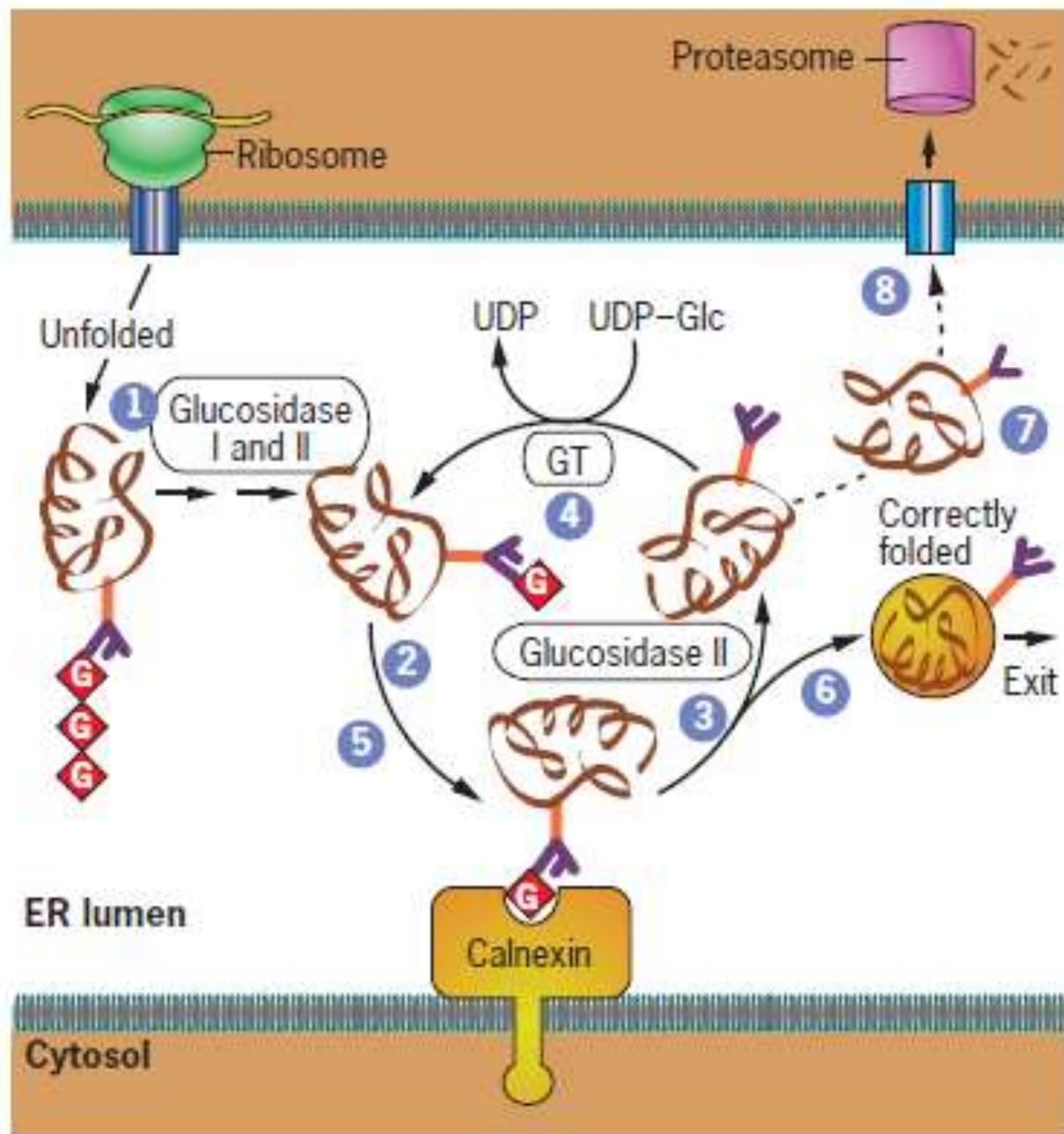


(a) Formation of a disulfide bond

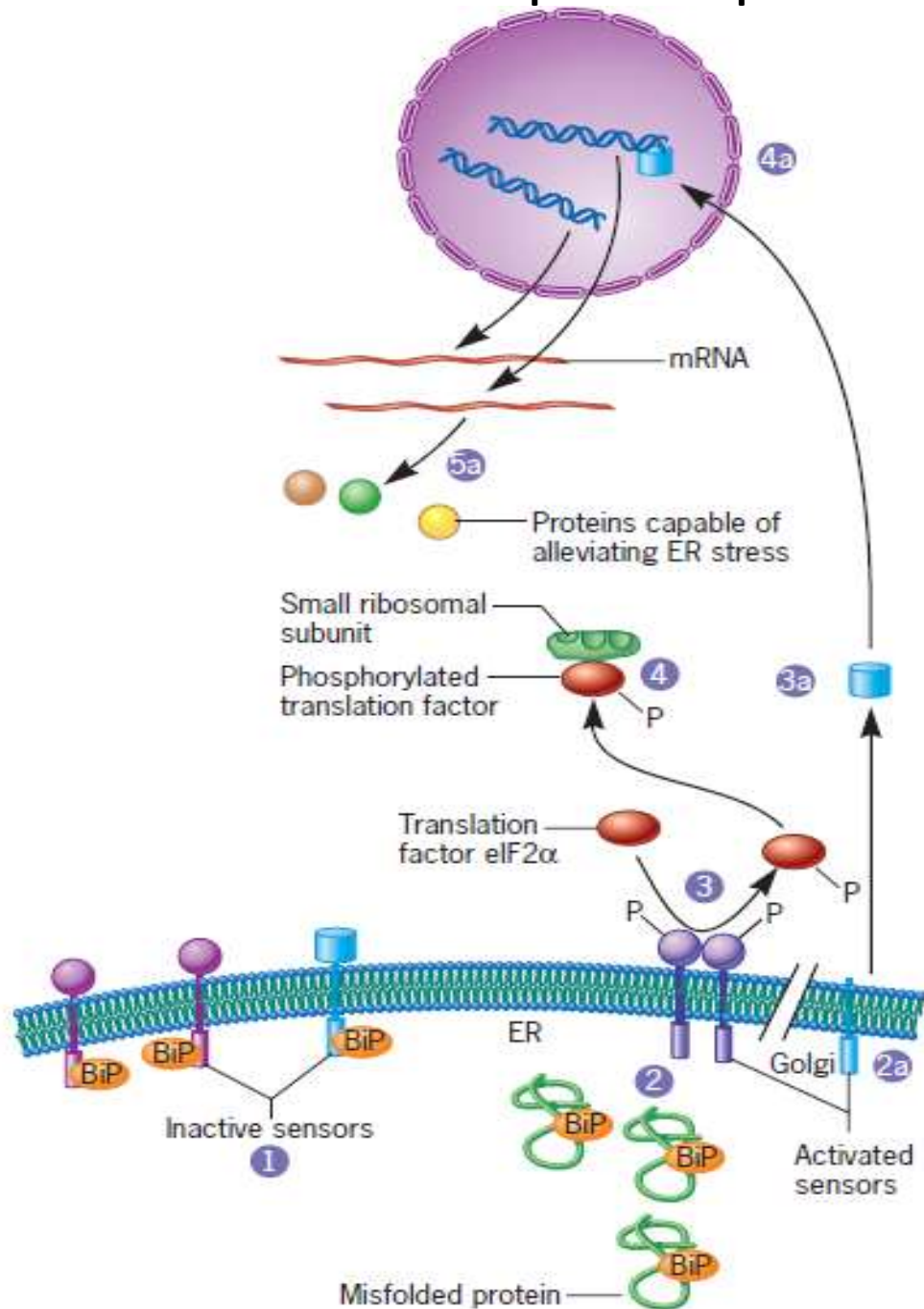


(b) Rearrangement of disulfide bonds





A model of the mammalian unfolded protein response



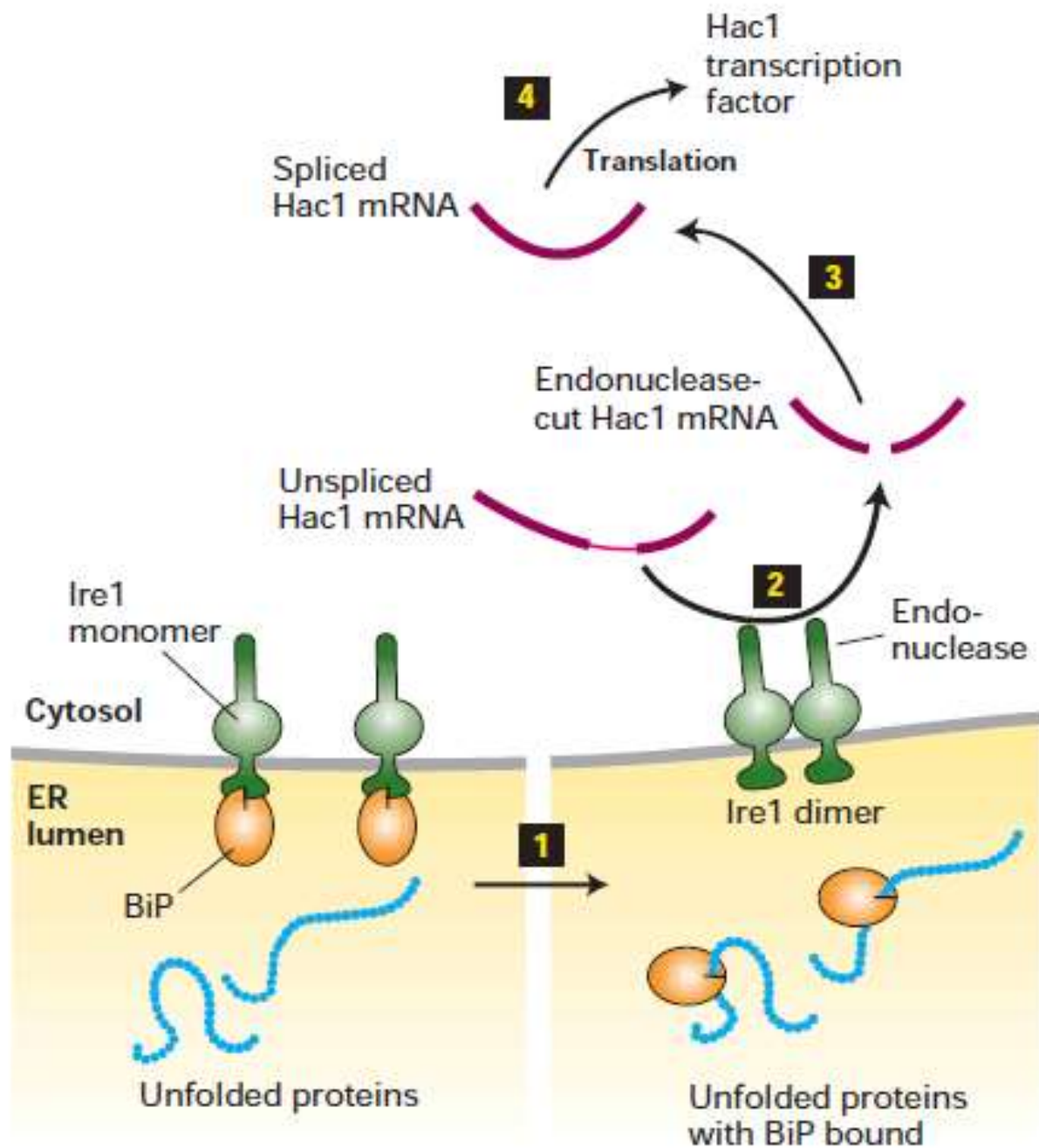
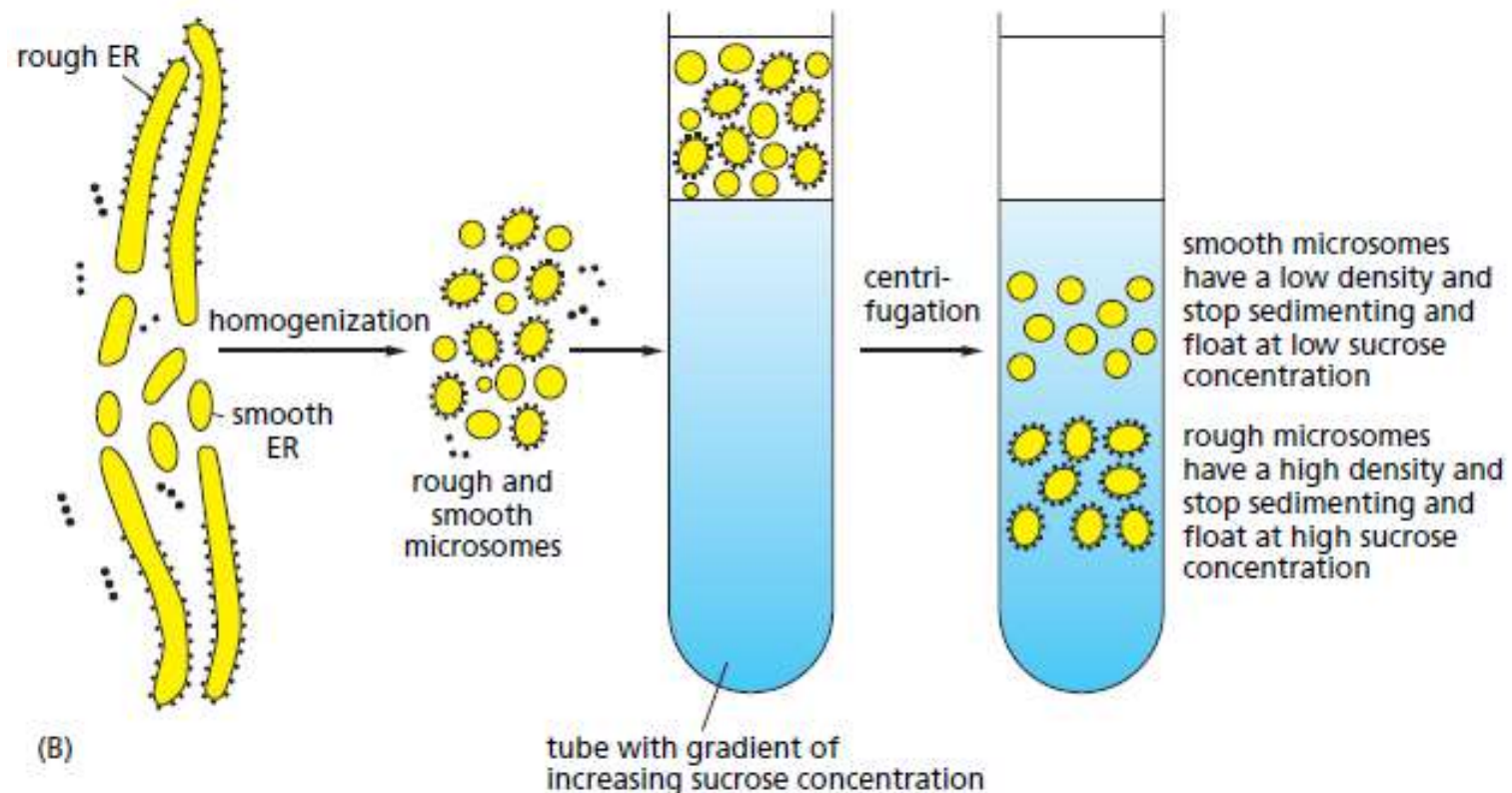
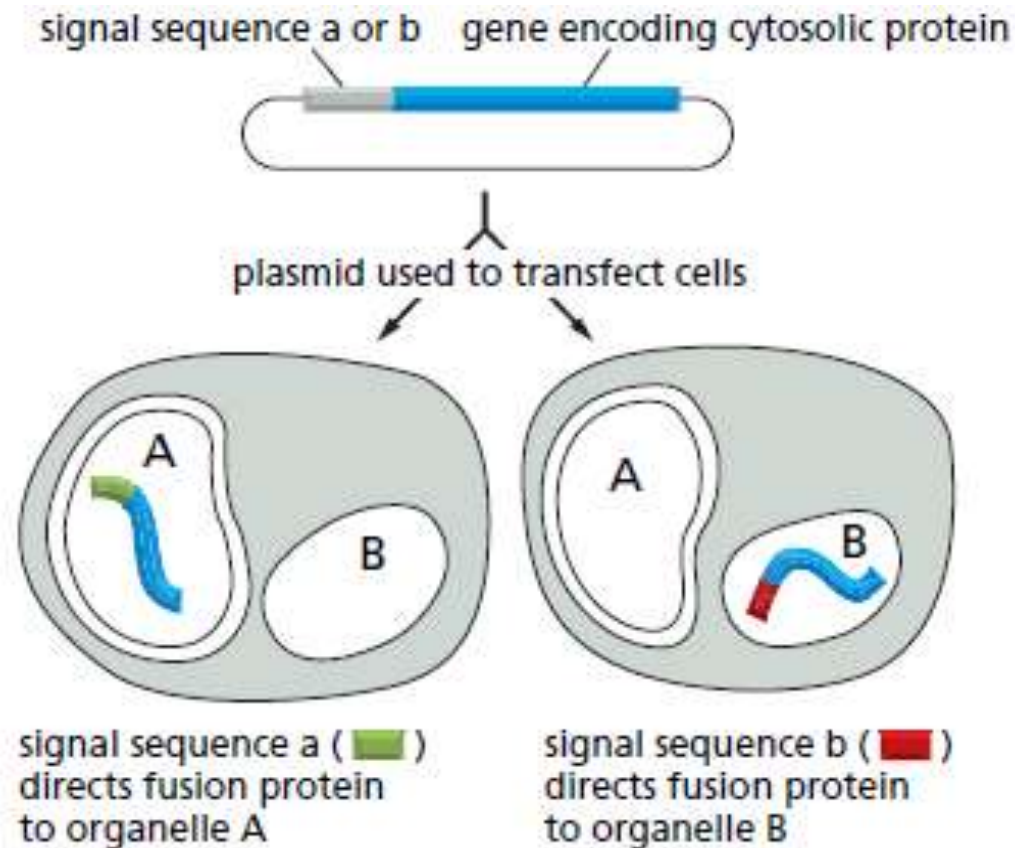


Table 12-3 Some Typical Signal Sequences

FUNCTION OF SIGNAL SEQUENCE	EXAMPLE OF SIGNAL SEQUENCE
Import into nucleus	-Pro-Pro-Lys-Lys-Lys-Arg-Lys-Val-
Export from nucleus	-Leu-Ala-Leu-Lys-Leu-Ala-Gly-Leu-Asp-Ile-
Import into mitochondria	⁺ H ₃ N-Met-Leu-Ser-Leu-Arg-Gln-Ser-Ile-Arg-Phe-Phe-Lys-Pro-Ala-Thr-Arg-Thr-Leu-Cys-Ser-Ser-Arg-Tyr-Leu-Leu-
Import into plastid	⁺ H ₃ N-Met-Val-Ala-Met-Ala-Met-Ala-Ser-Leu-Gln-Ser-Ser-Met-Ser-Ser-Leu-Ser-Leu-Ser-Ser-Asn-Ser-Phe-Leu-Gly-Gln-Pro-Leu-Ser-Pro-Ile-Thr-Leu-Ser-Pro-Phe-Leu-Gln-Gly-
Import into peroxisomes	-Ser-Lys-Leu-COO ⁻
Import into ER	⁺ H ₃ N-Met-Met-Ser-Phe-Val-Ser-Leu-Leu-Leu-Val-Gly-Ile-Leu-Phe-Trp-Ala-Thr-Glu-Ala-Glu-Gln-Leu-Thr-Lys-Cys-Glu-Val-Phe-Gln-
Return to ER	-Lys-Asp-Glu-Leu-COO ⁻



A TRANSFECTION APPROACH FOR DEFINING SIGNAL SEQUENCES



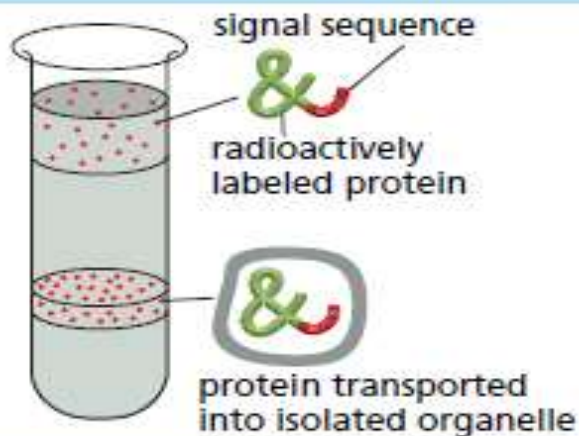
By altering the signal sequence using site-directed mutagenesis, we can determine which structural features are important for its function.

A BIOCHEMICAL APPROACH FOR STUDYING THE MECHANISM OF PROTEIN TRANSLOCATION

In this approach, a labeled protein containing a specific signal sequence is transported into isolated organelles *in vitro*. The labeled protein is usually produced by cell-free translation of a purified mRNA encoding the protein. Radioactive amino acids are used to label the newly synthesized protein so that it can be distinguished from the many other proteins that are present in the *in vitro* translation system.

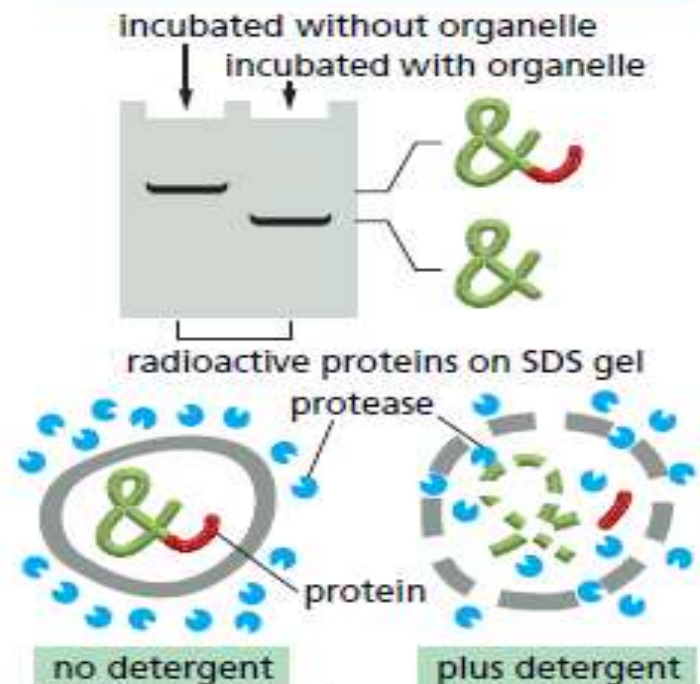
Three methods are commonly used to test if the labeled protein has been translocated into the organelle:

1. The labeled protein co-fractionates with the organelle during centrifugation.



3. The protein is protected from digestion when proteases are added to the incubation medium, but is susceptible if a detergent is first added to disrupt the organelle membrane.

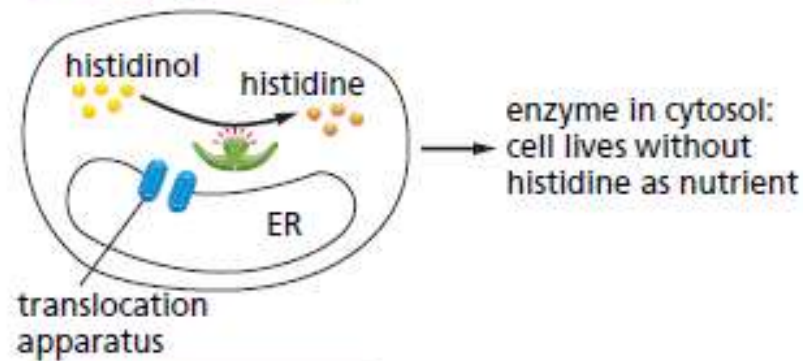
2. The signal sequence is removed by a specific protease that is present inside the organelle.



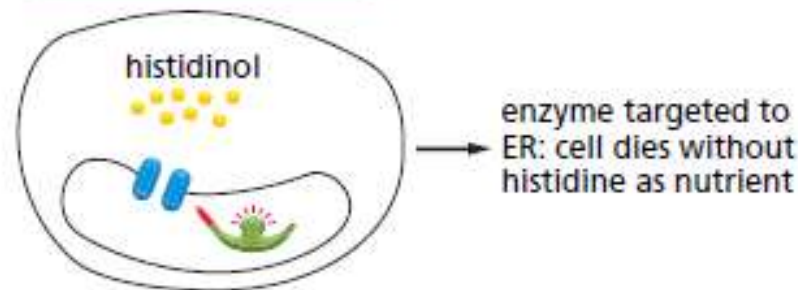
By exploiting such *in vitro* assays, one can determine what components (proteins, ATP, GTP, etc.) are required for the translocation process.

GENETIC APPROACHES FOR STUDYING THE MECHANISM OF PROTEIN TRANSLOCATION

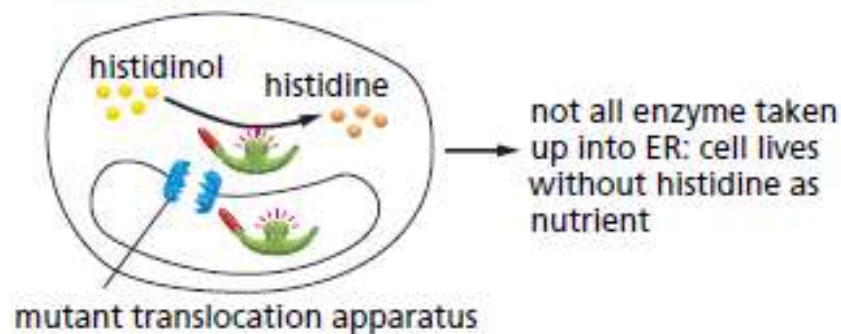
wild-type yeast cell

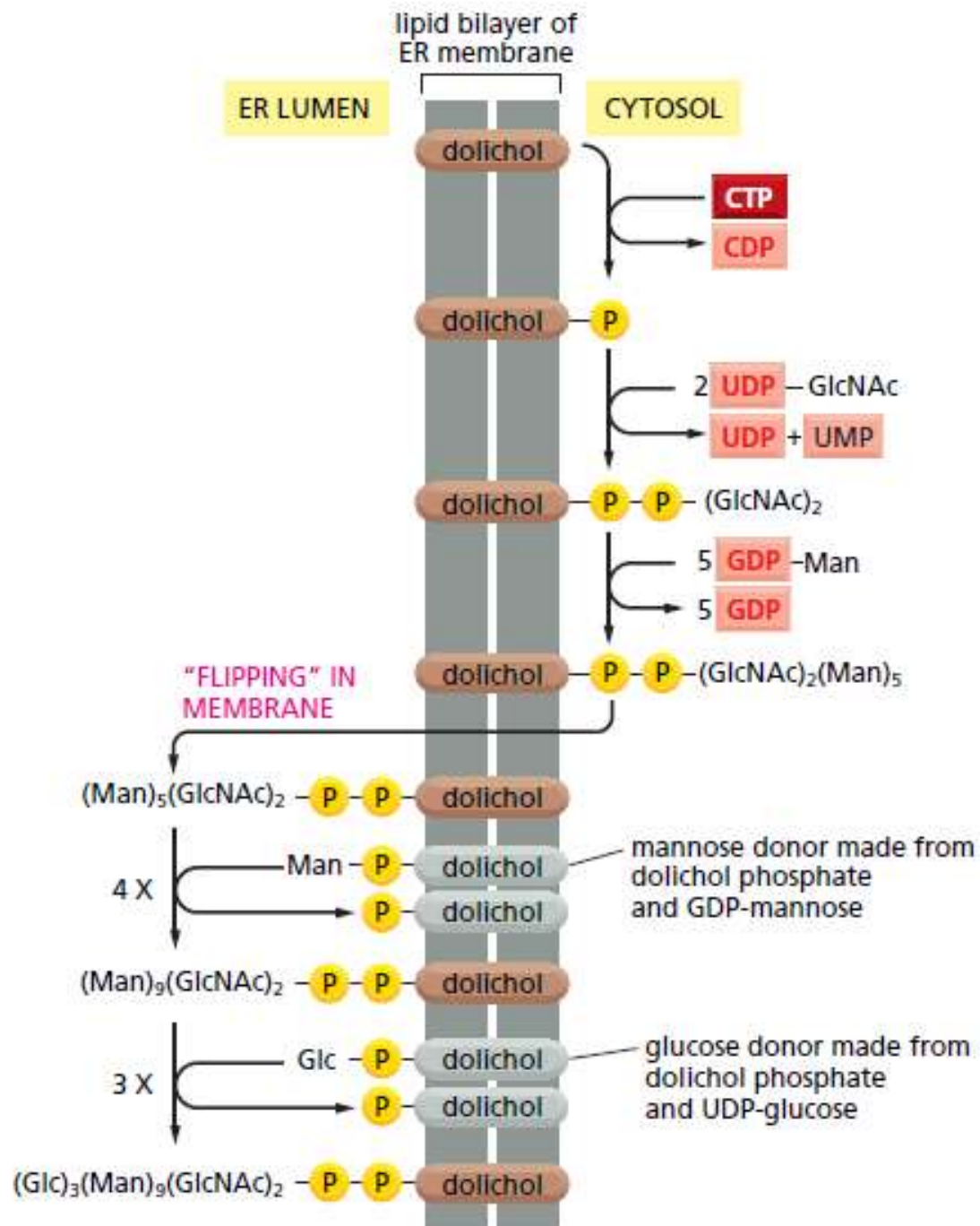


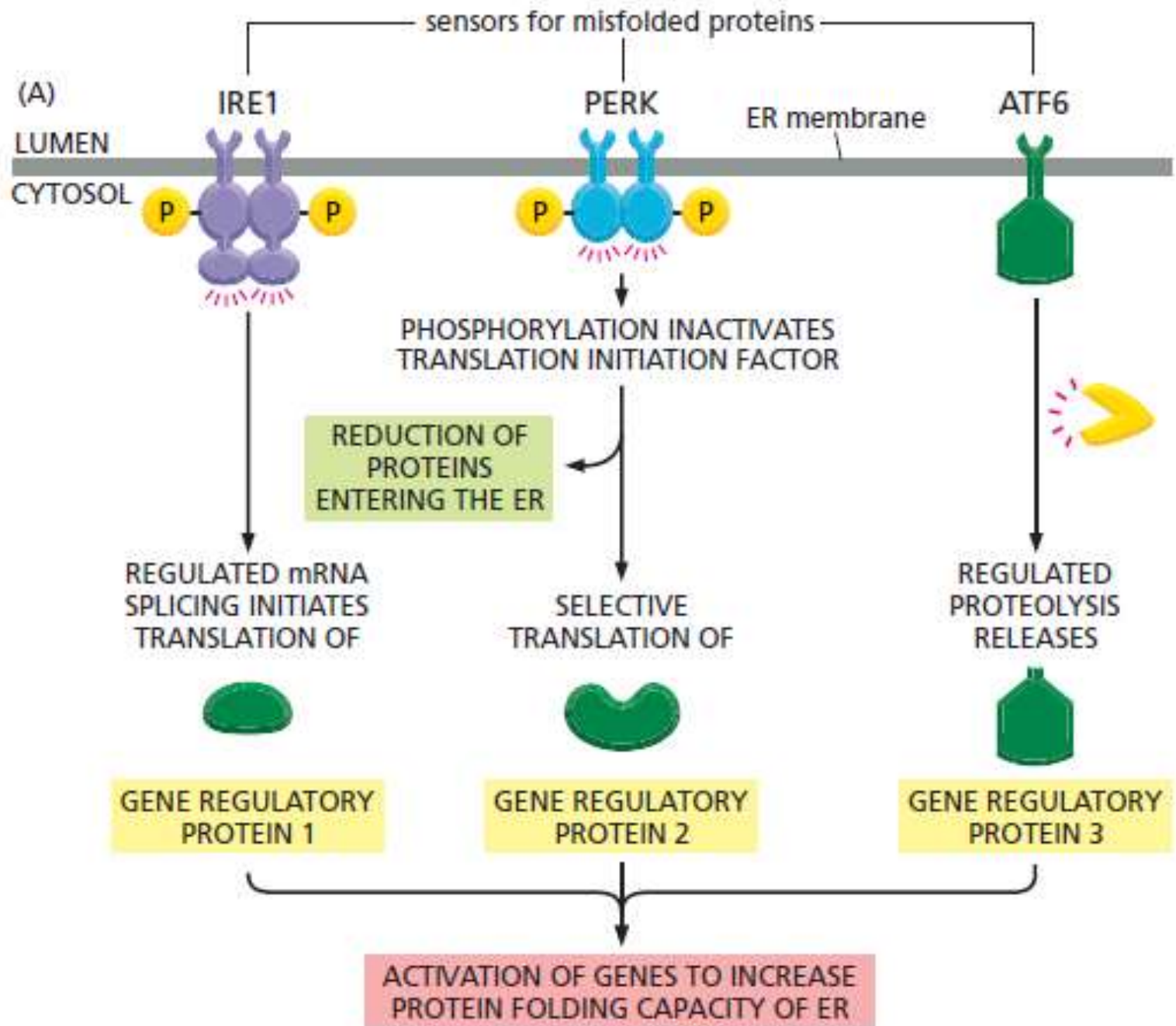
engineered yeast cell



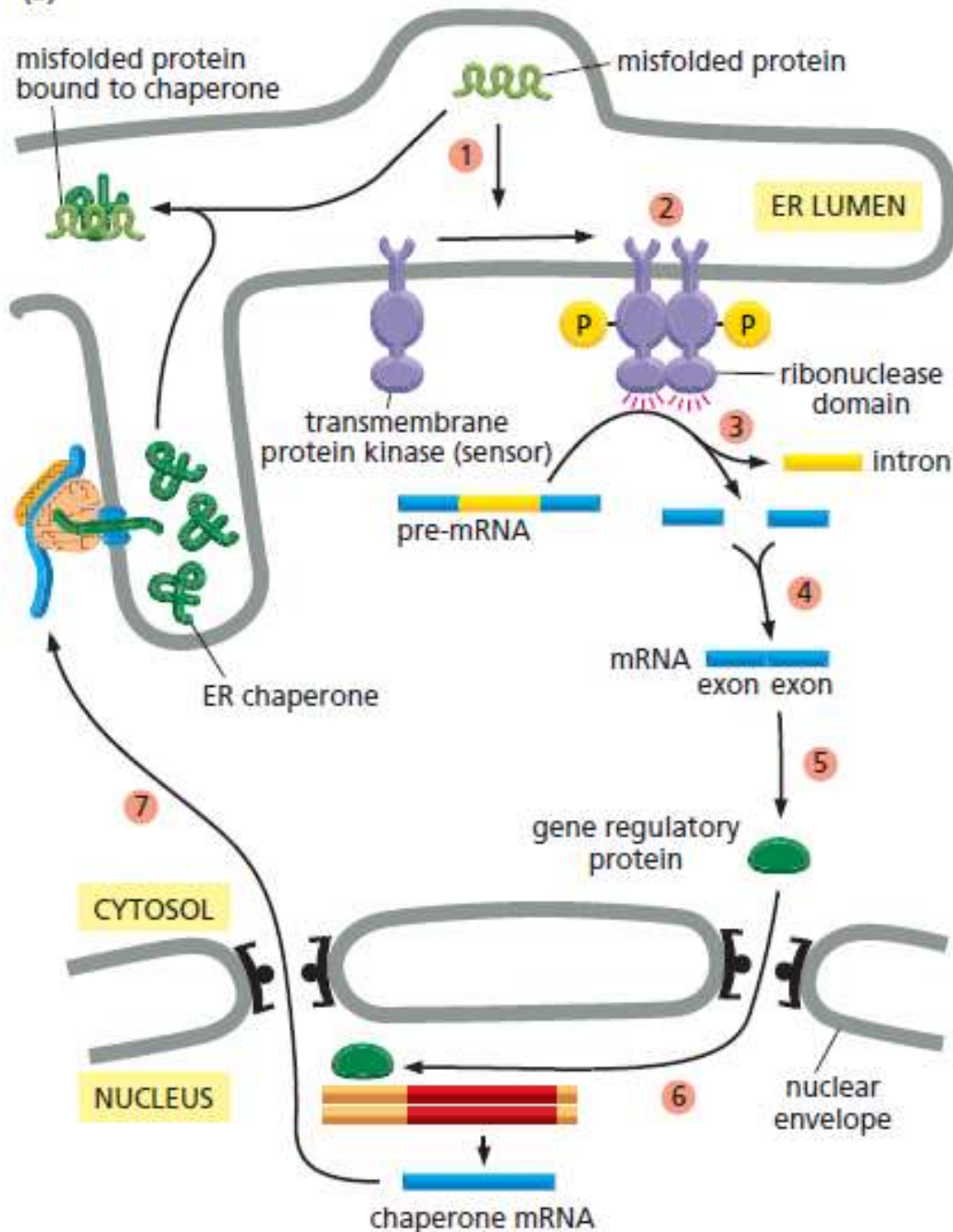
mutant engineered cell







(B)



1 MISFOLDED PROTEINS IN ER SIGNAL THE NEED FOR MORE ER CHAPERONES BY ACTIVATING A TRANSMEMBRANE KINASE

2 ACTIVATED KINASE TURNS INTO AN ENDORIBONUCLEASE

3 ENDORIBONUCLEASE CUTS SPECIFIC RNA MOLECULES AT TWO POSITIONS, REMOVING AN INTRON

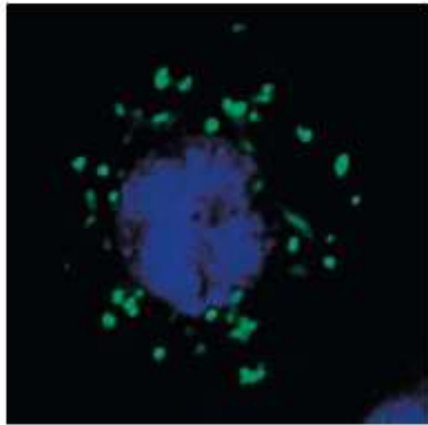
4 TWO EXONS ARE LIGATED TO FORM AN ACTIVE mRNA

5 mRNA IS TRANSLATED TO MAKE A GENE REGULATORY PROTEIN

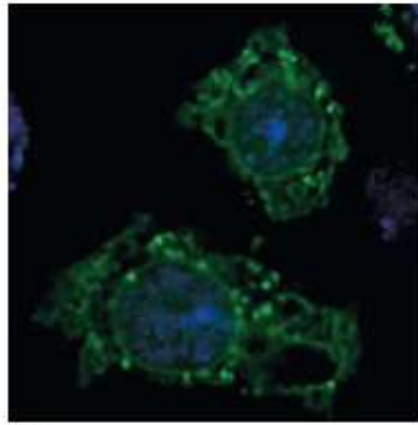
6 GENE REGULATORY PROTEIN ENTERS NUCLEUS AND ACTIVATES GENES ENCODING ER CHAPERONES

7 CHAPERONES ARE MADE IN ER, WHERE THEY HELP FOLD PROTEINS

Inhibition of gene expression with RNA interference

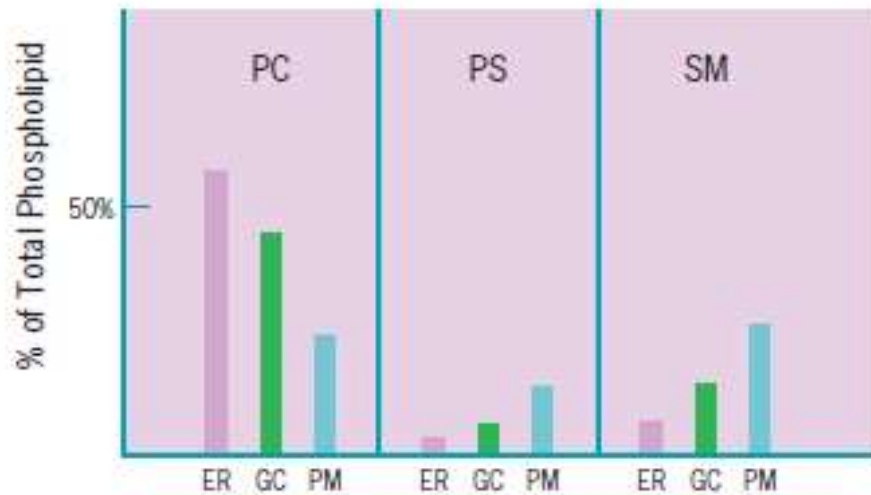


(a)



(b)

Maintenance of membrane asymmetry.



ER = Endoplasmic reticulum

GC = Golgi complex

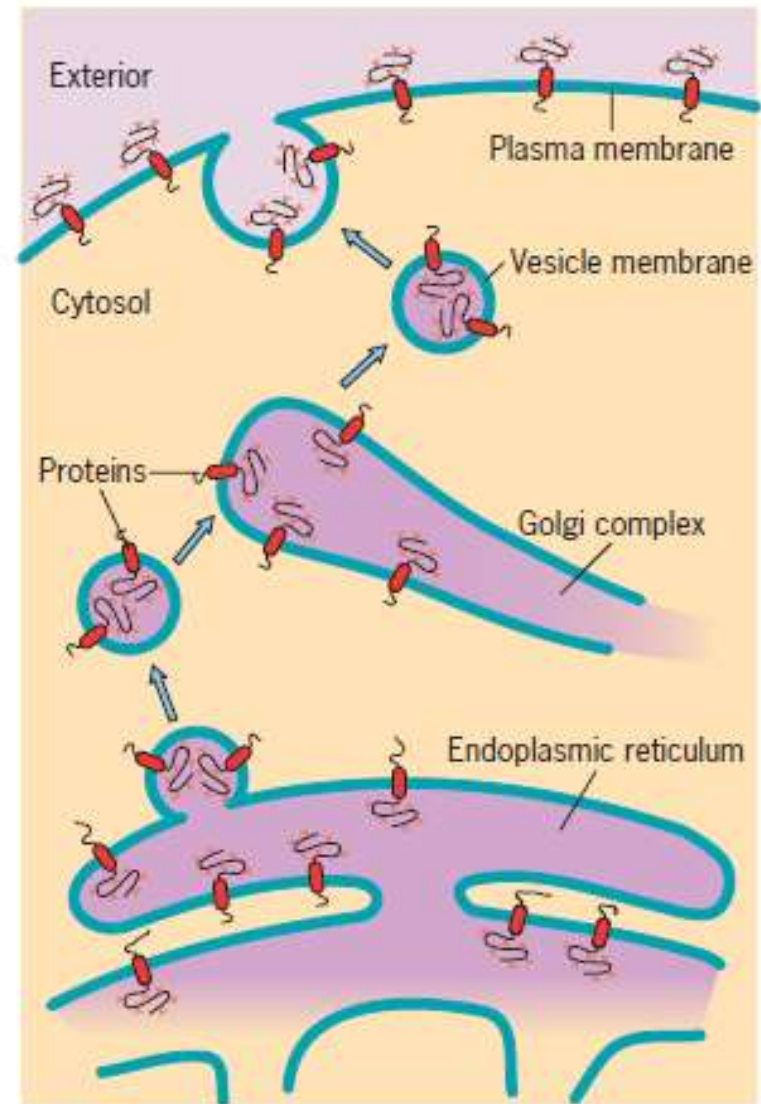
PM = Erythrocyte plasma membrane

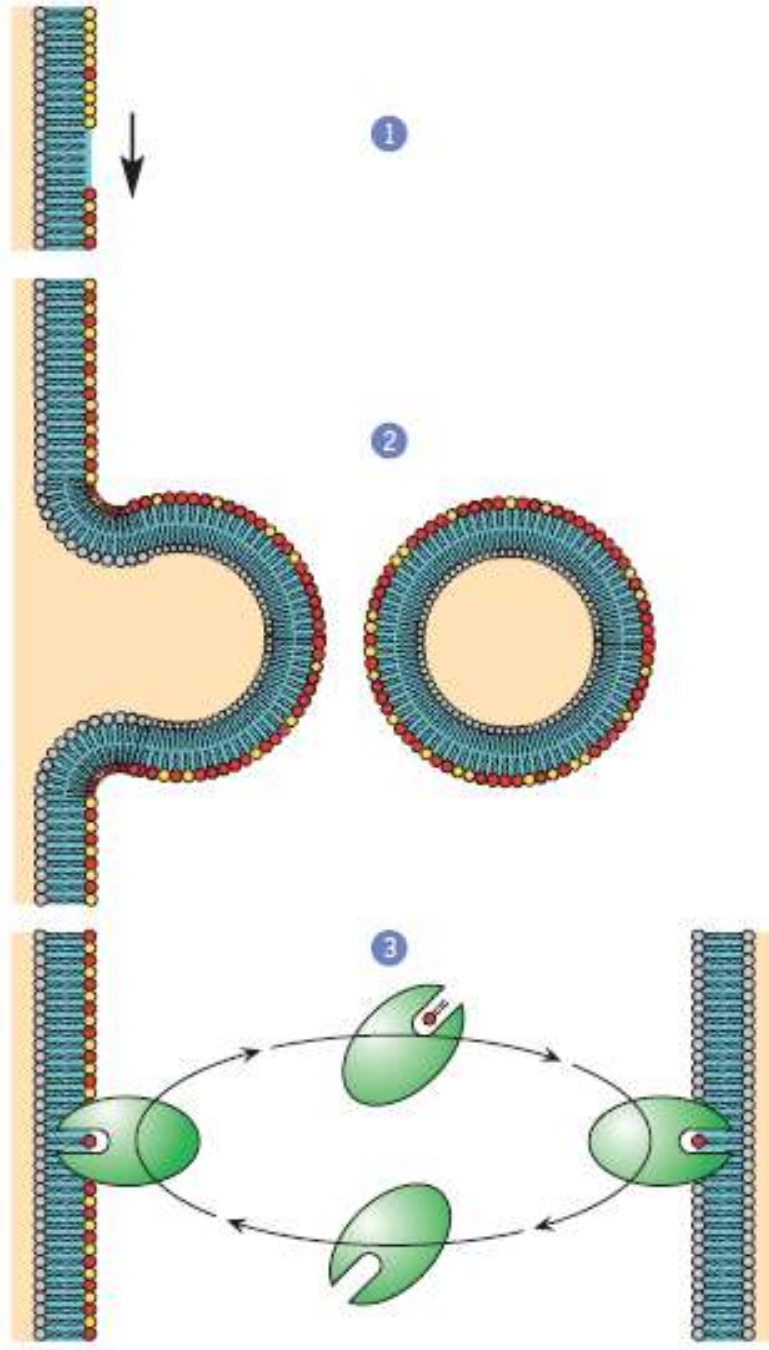
PC = Phosphatidylcholine

PS = Phosphatidylserine

SM = Sphingomyelin

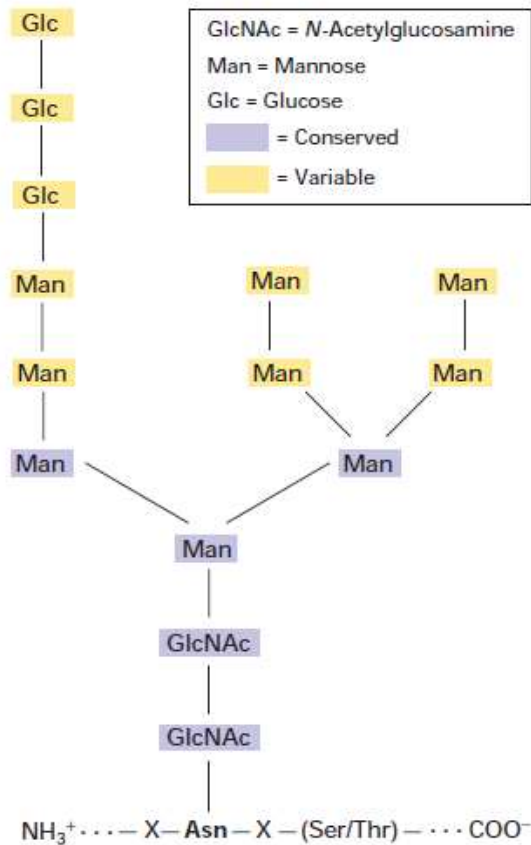
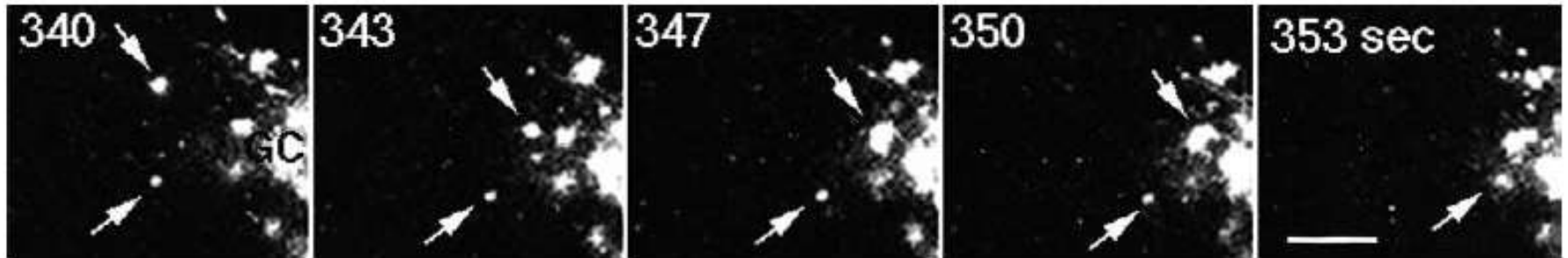
(a)

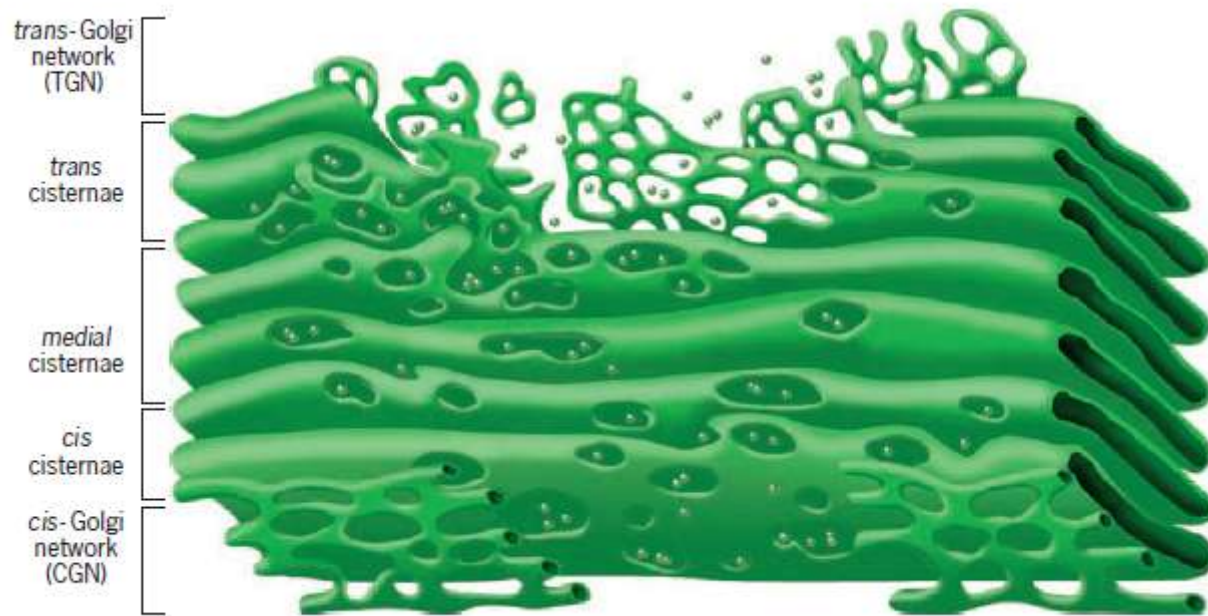




(b)

Visualizing membrane traffic with the use of a fluorescent tag



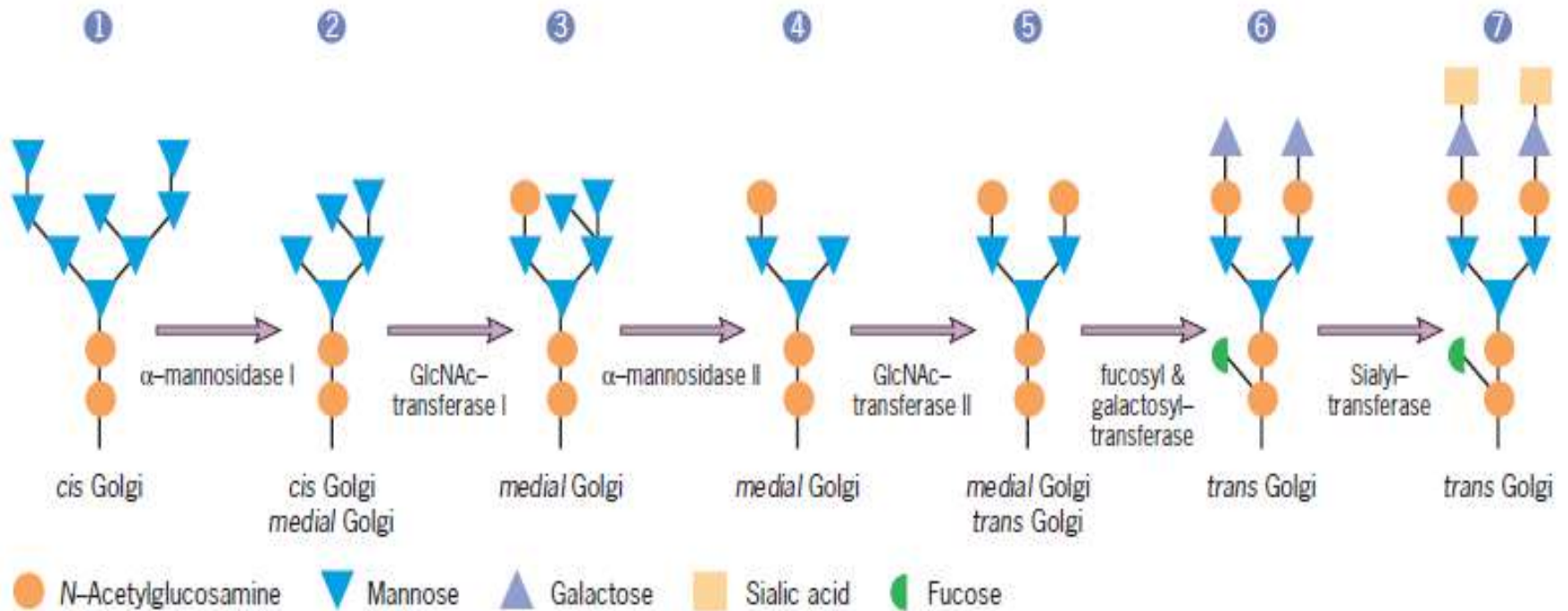


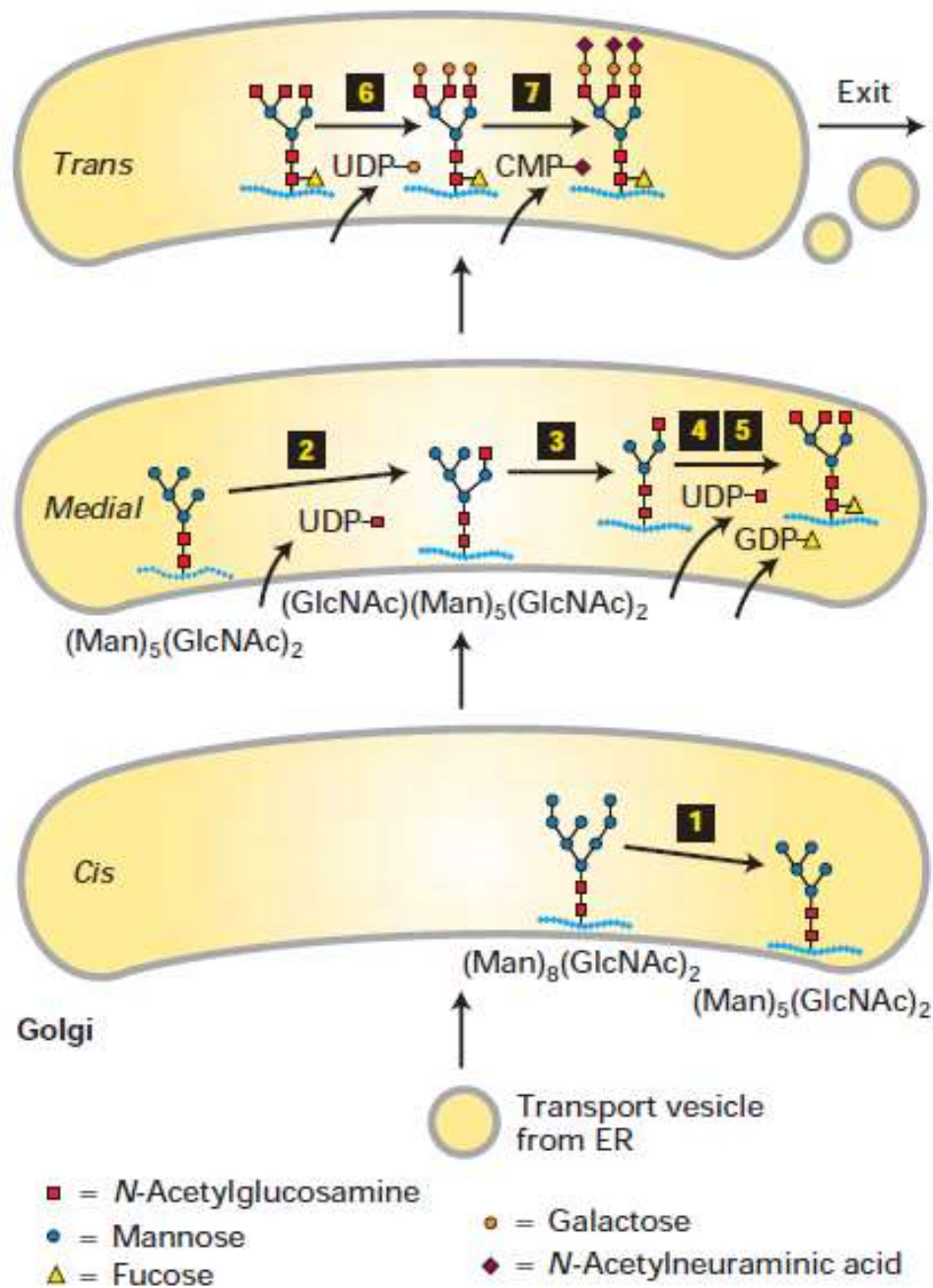
(a)



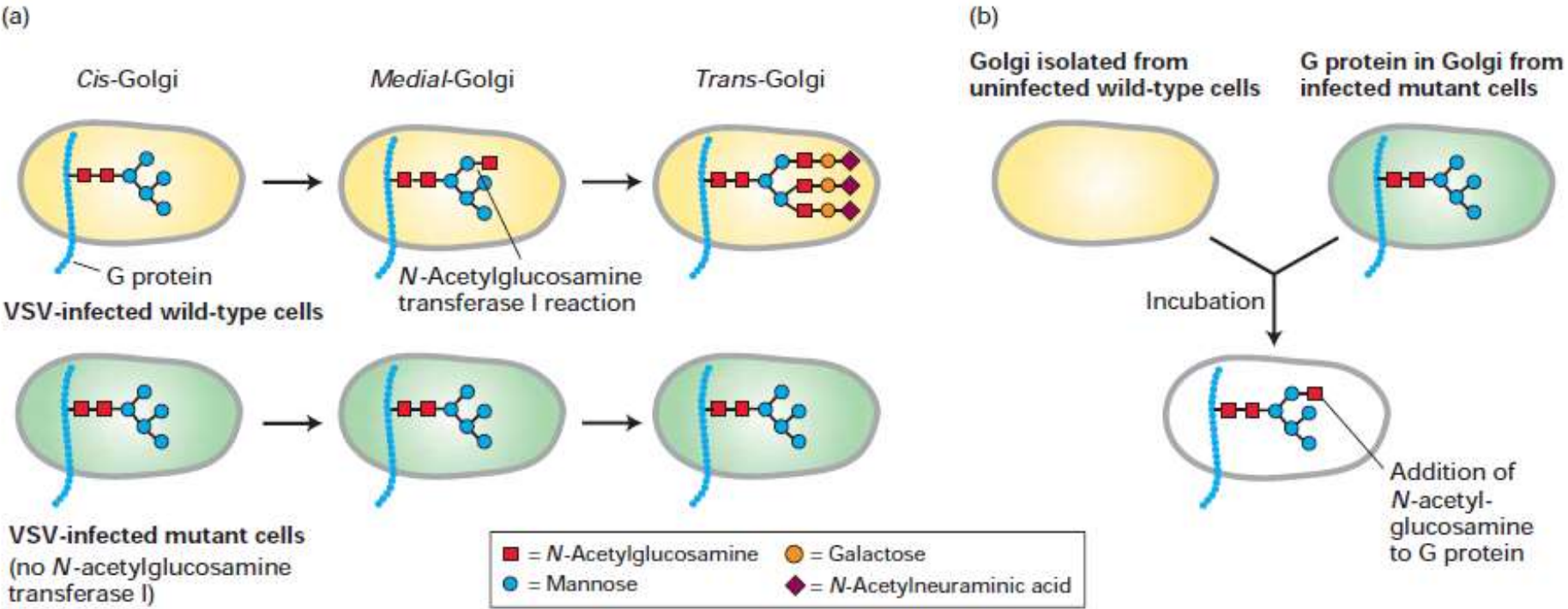
(b)

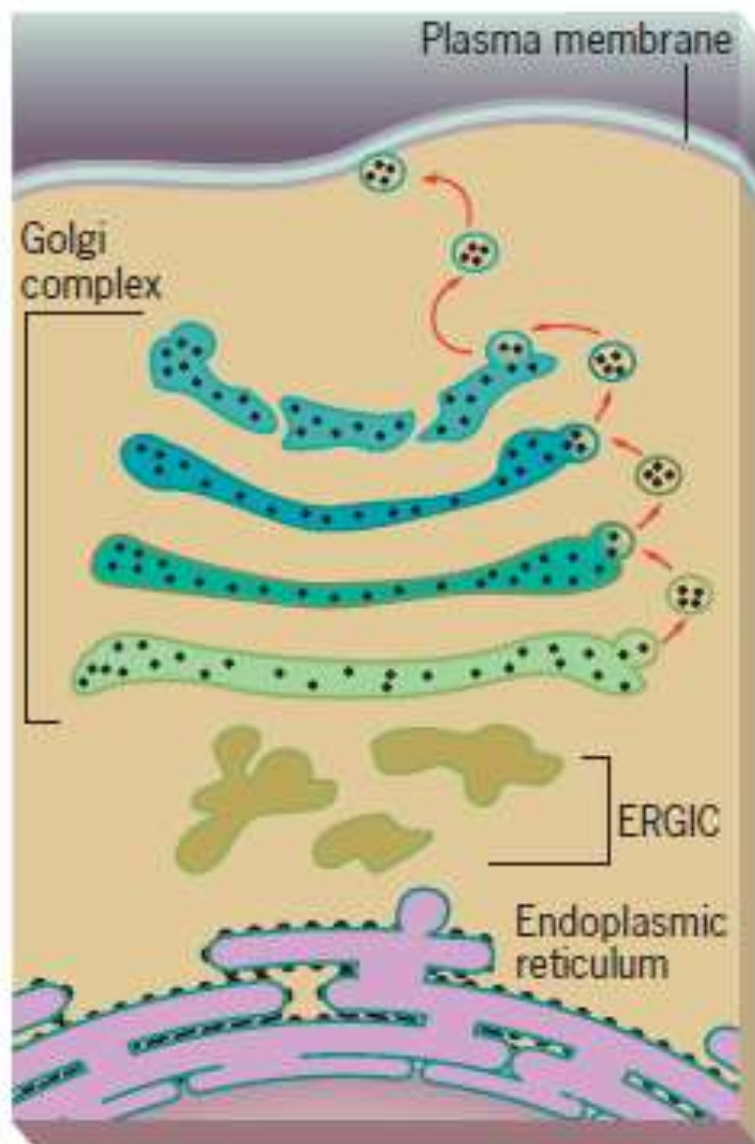
Steps in the glycosylation of a typical mammalian *N-linked oligosaccharide in the Golgi complex*



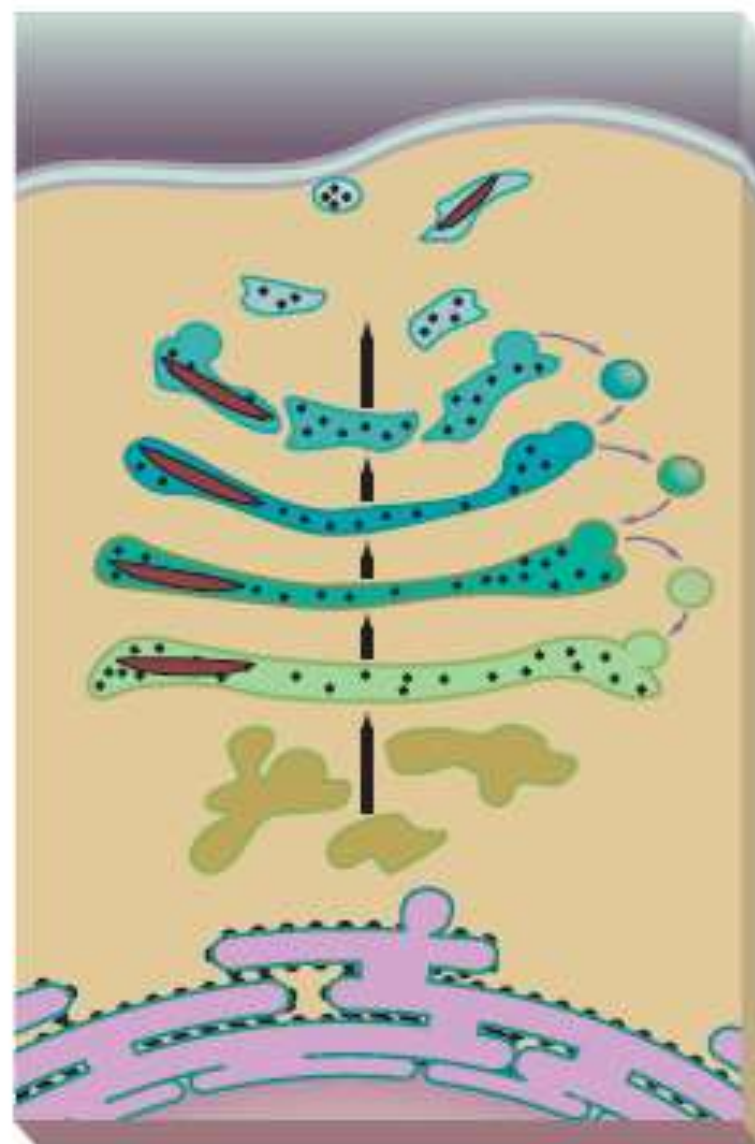


Protein transport from one Golgi cisternae to another can be assayed in a cell-free system

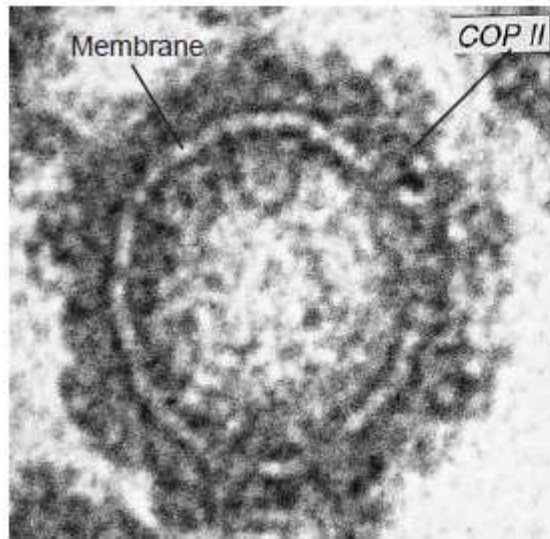
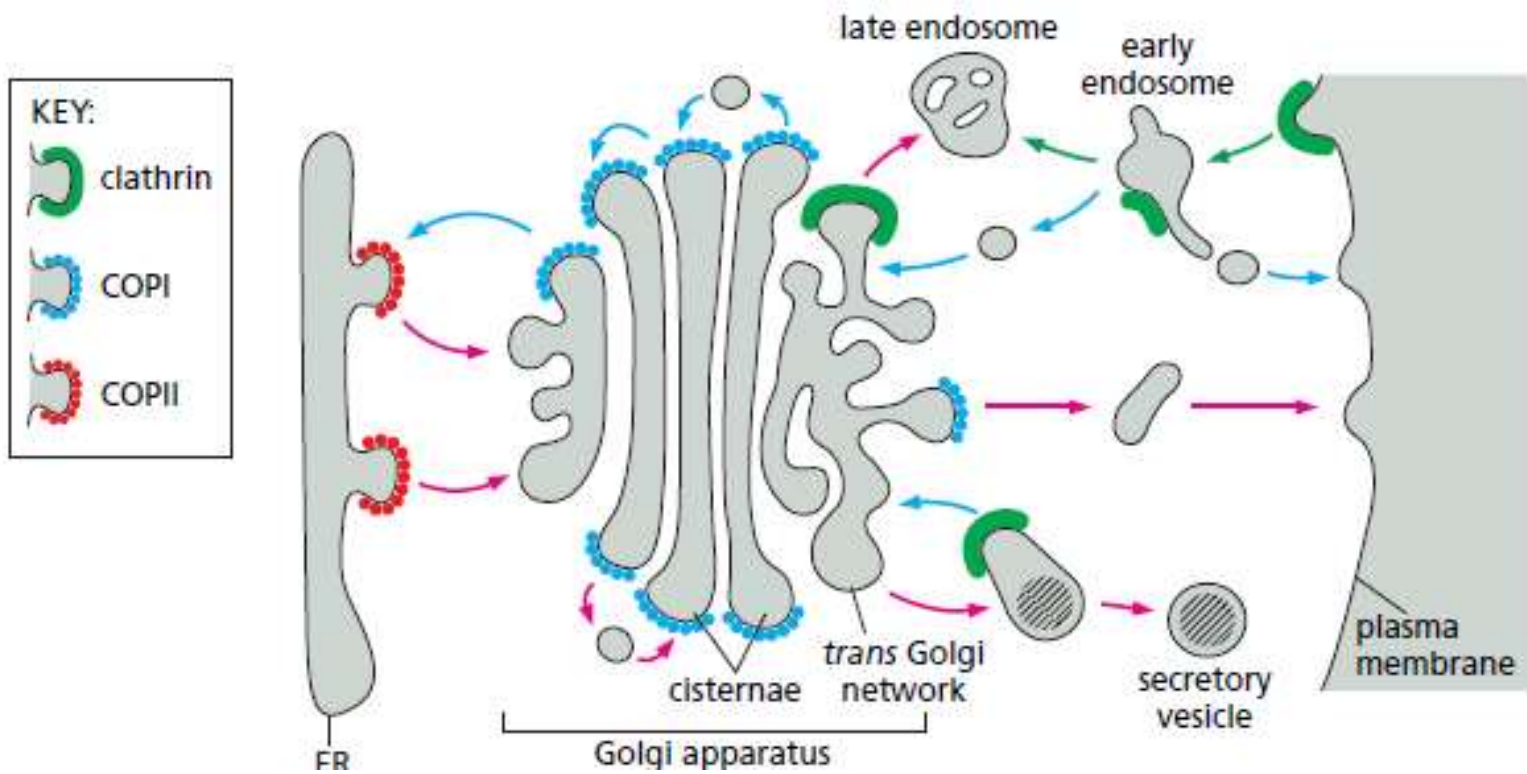




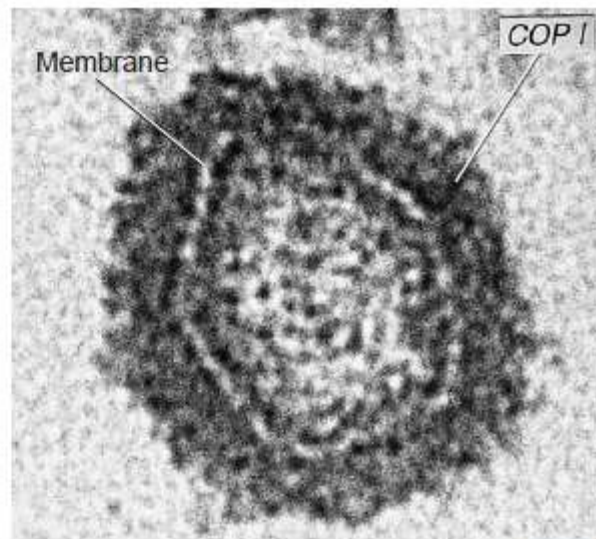
(a) Vesicular transport model



(b) Cisternal maturation model

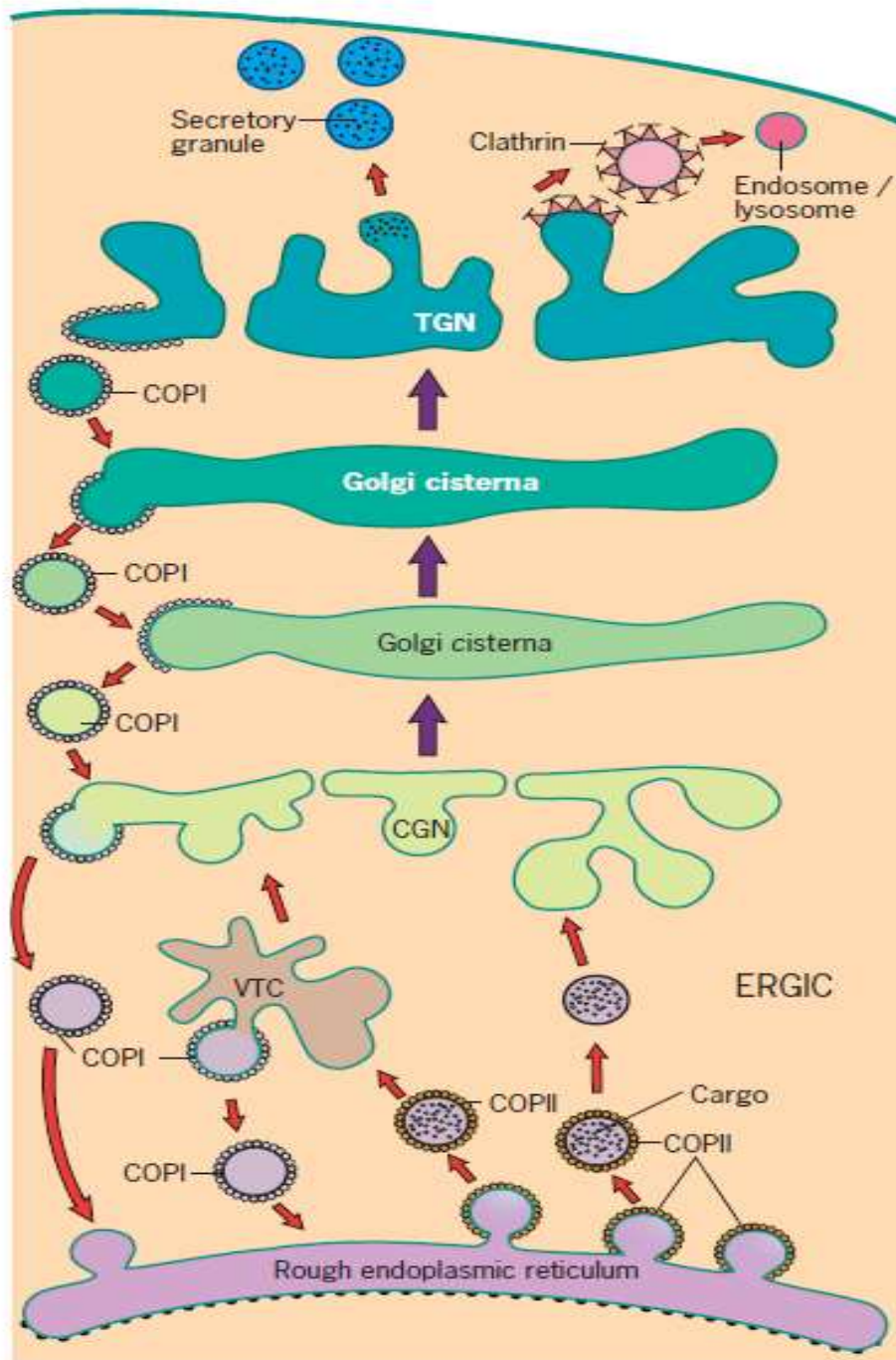


(a)

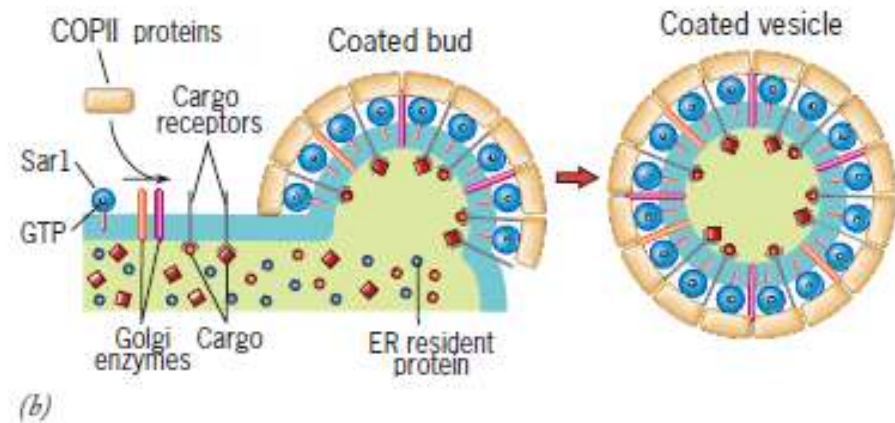


(b)

20 nm

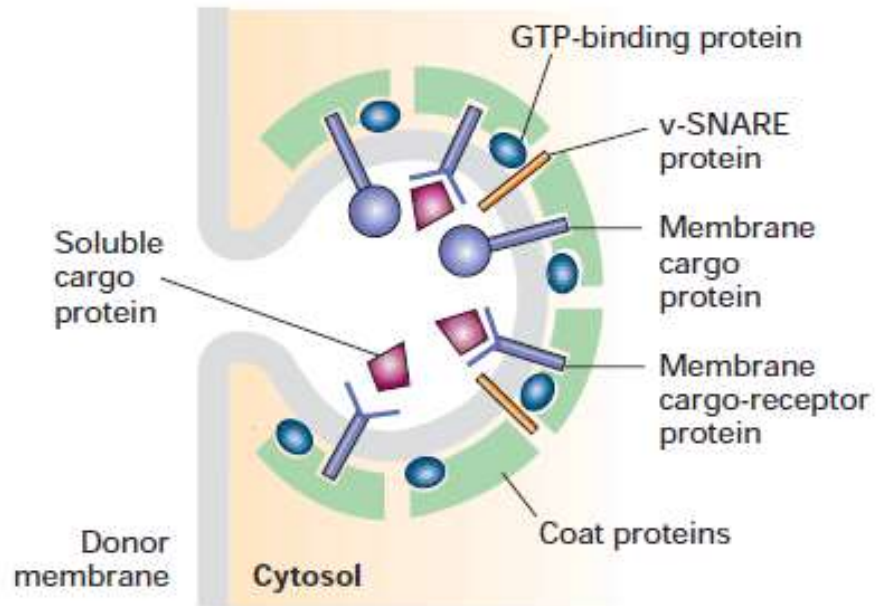


(a)



(b)

(a) Coated vesicle budding



(b) Uncoated vesicle fusion

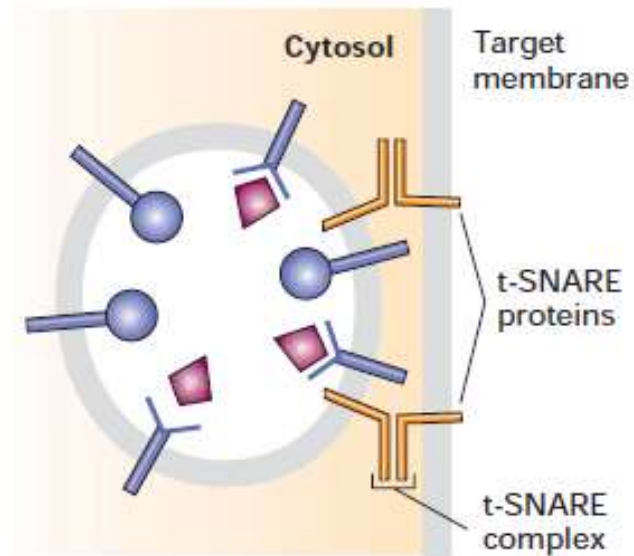
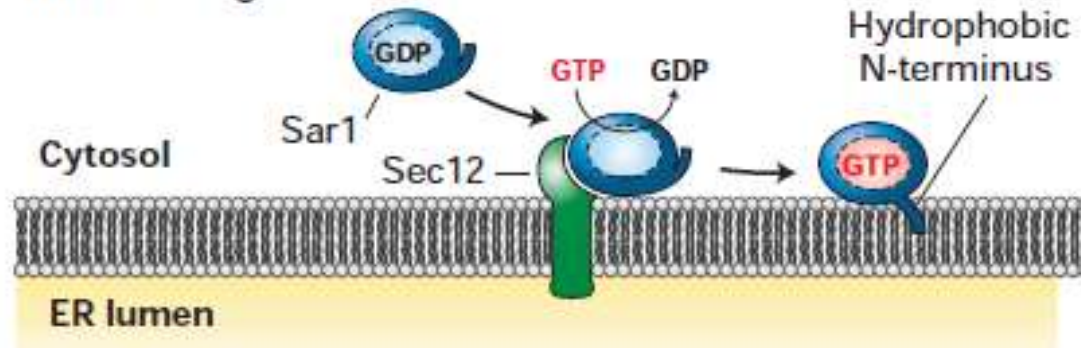


TABLE 17-1 Coated Vesicles Involved in Protein Trafficking

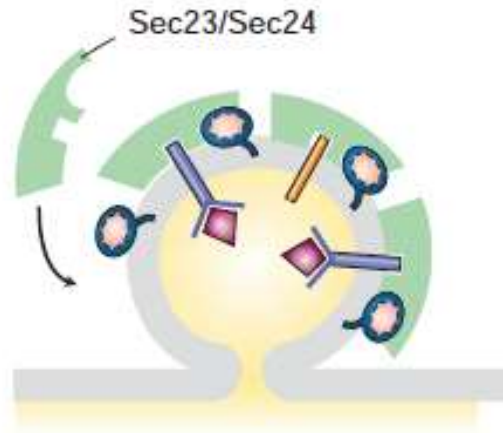
Vesicle Type	Coat Proteins	Associated GTPase	Transport Step Mediated
COPII	Sec23/Sec24 and Sec13/Sec31 complexes, Sec16	Sar1	ER to <i>cis</i> -Golgi
COPI	Coatomers containing seven different COP subunits	ARF	<i>cis</i> -Golgi to ER Later to earlier Golgi cisternae
Clathrin and adapter proteins*	Clathrin + AP1 complexes	ARF	<i>trans</i> -Golgi to endosome
	Clathrin + GGA	ARF	<i>trans</i> -Golgi to endosome
	Clathrin + AP2 complexes	ARF	Plasma membrane to endosome
	AP3 complexes	ARF	Golgi to lysosome, melanosome, or platelet vesicles

*Each type of AP complex consists of four different subunits. It is not known whether the coat of AP3 vesicles contains clathrin.

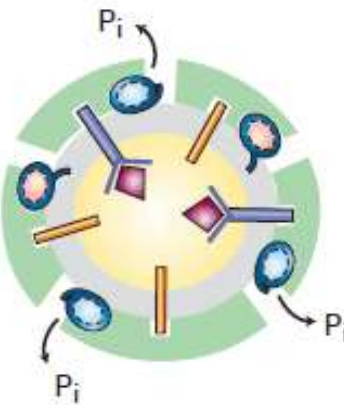
1 Sar1 membrane binding, GTP exchange



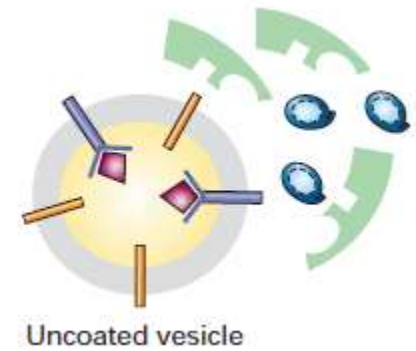
2 COPII coat assembly

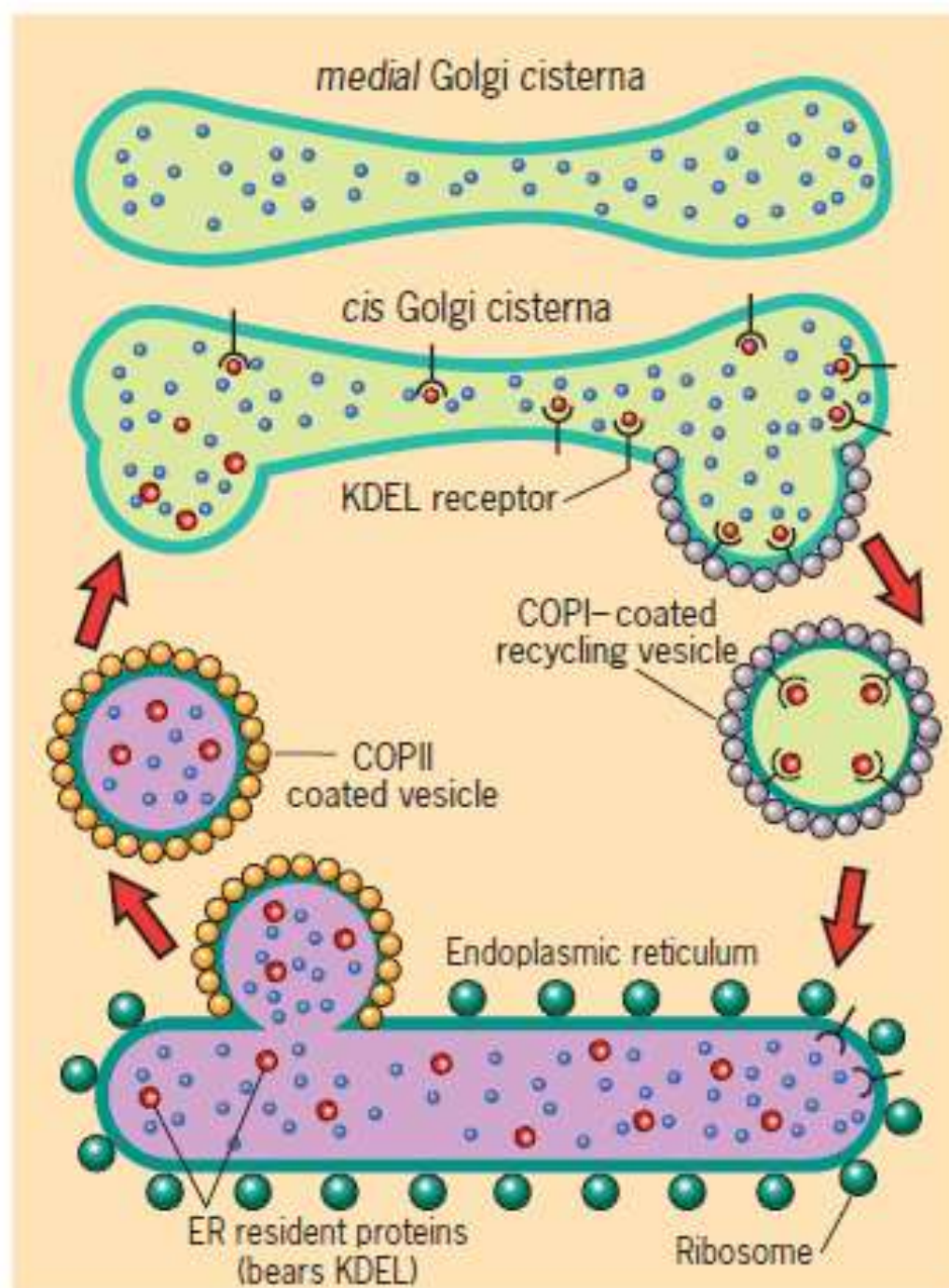


3 GTP hydrolysis

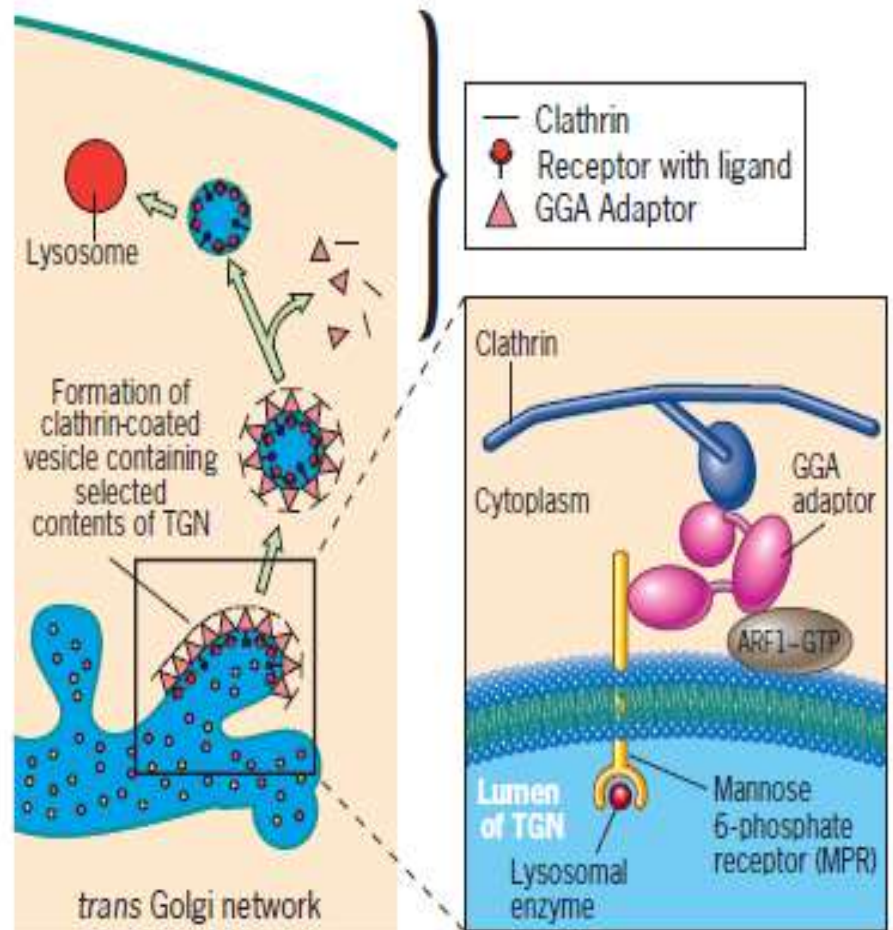
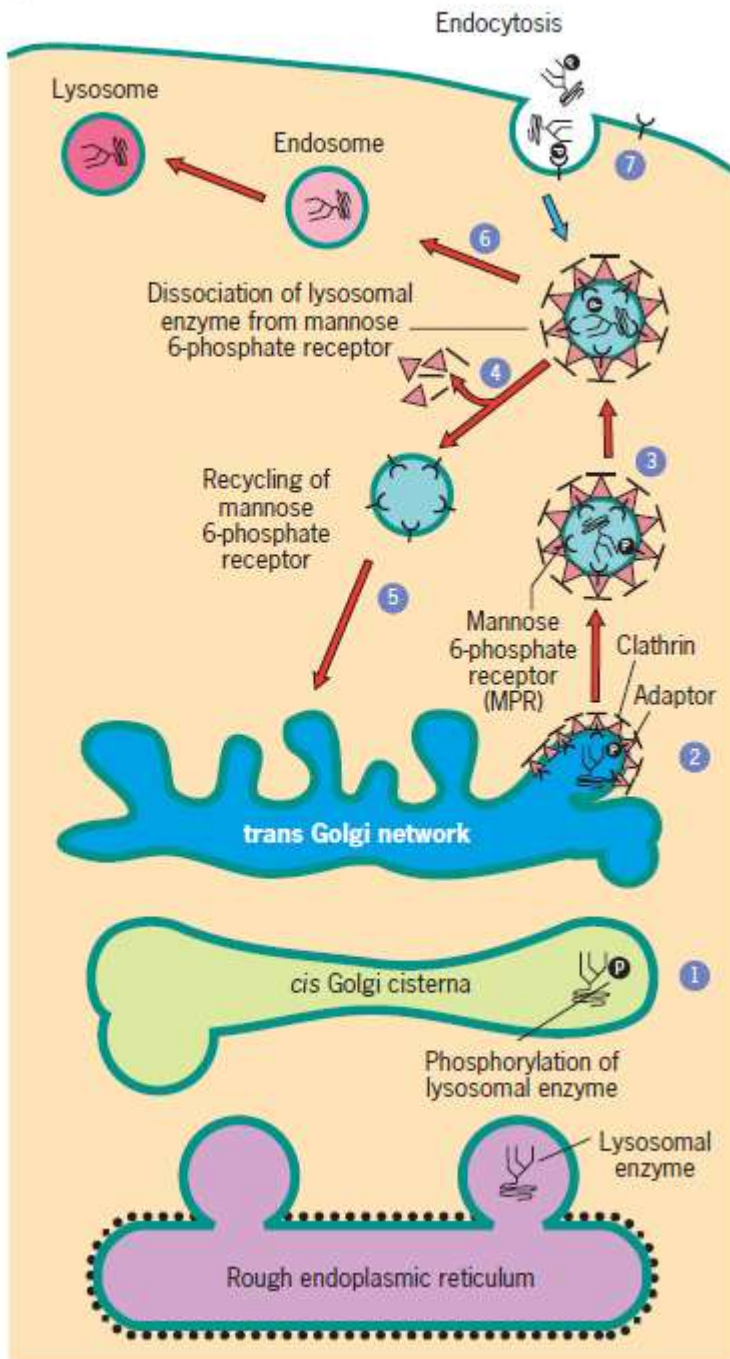


4 Coat disassembly

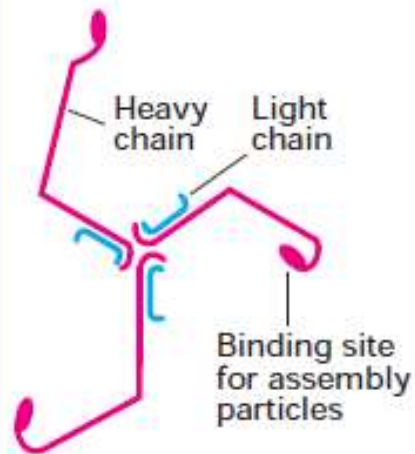




(a)



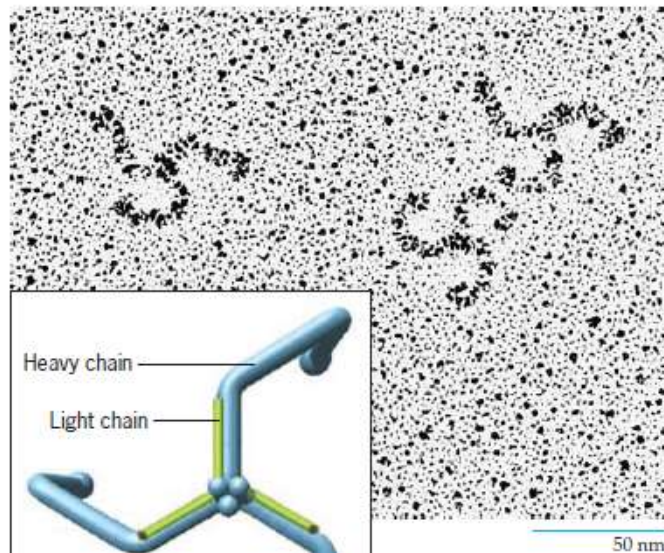
(a) Triskelion structure



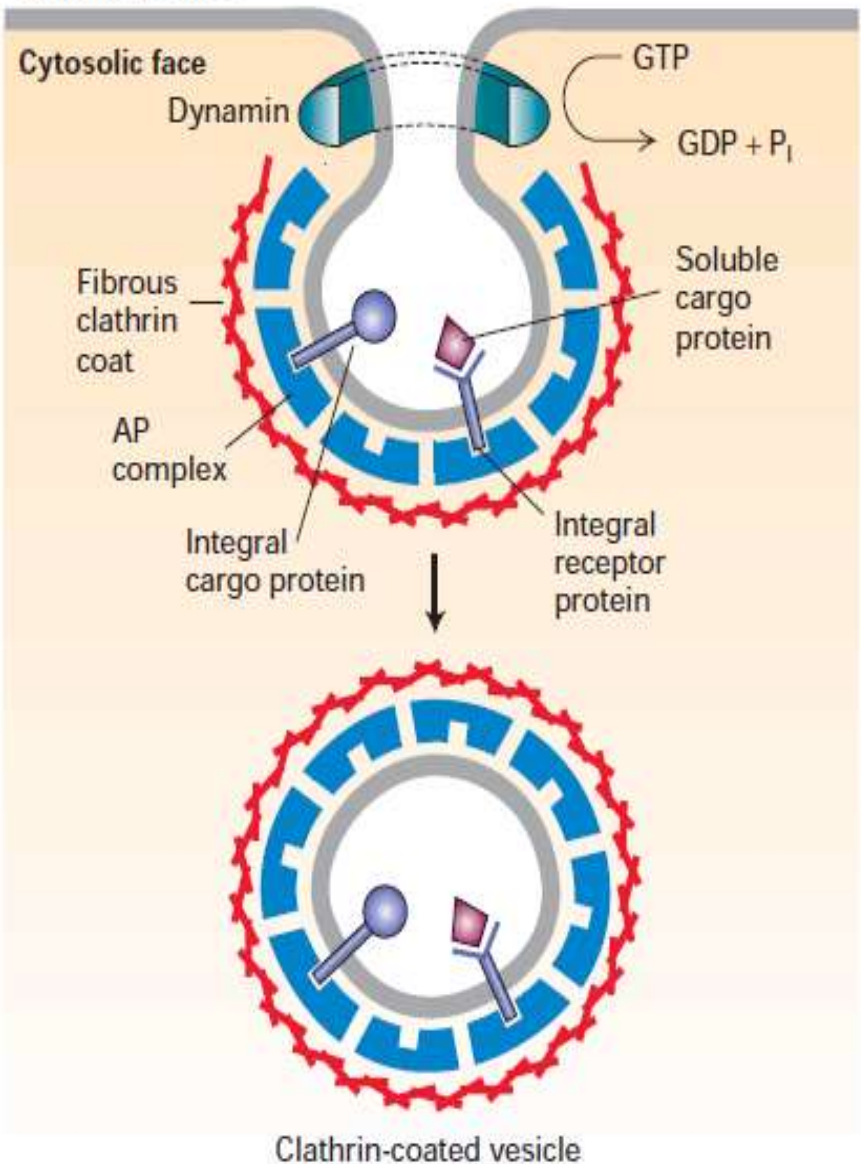
(b) Assembly intermediate

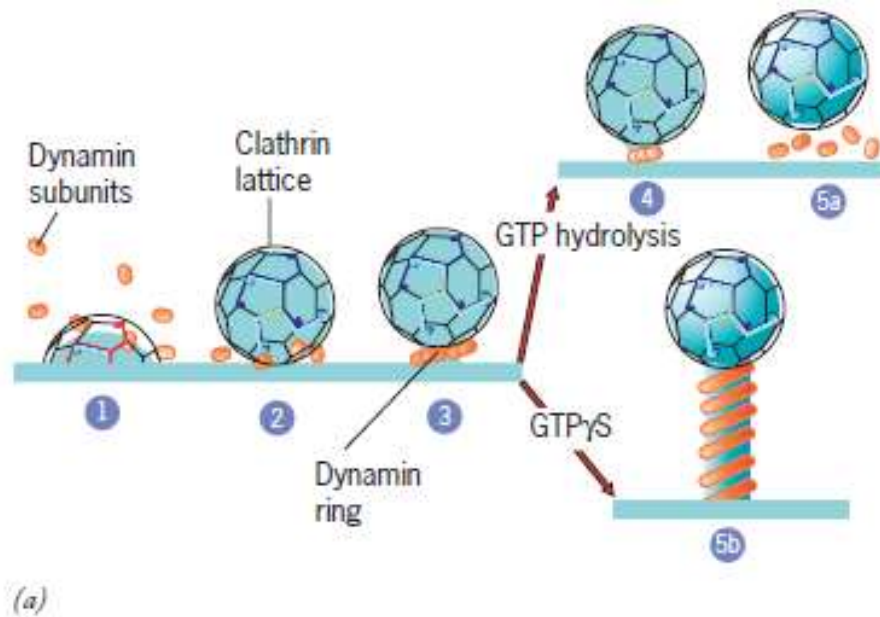
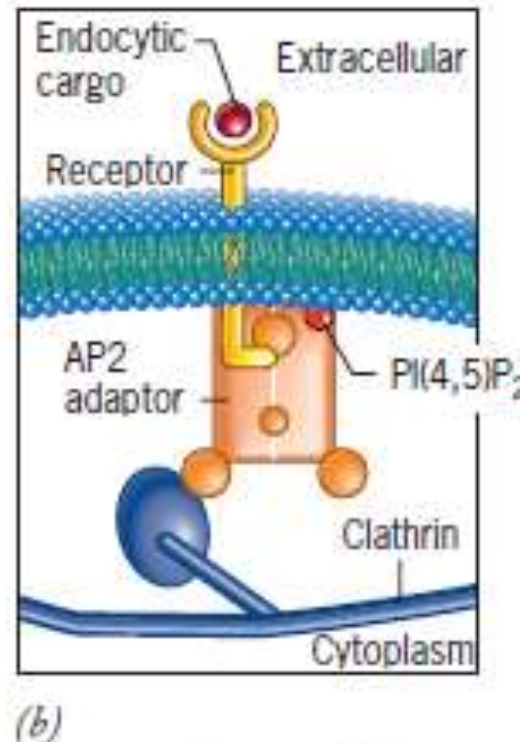
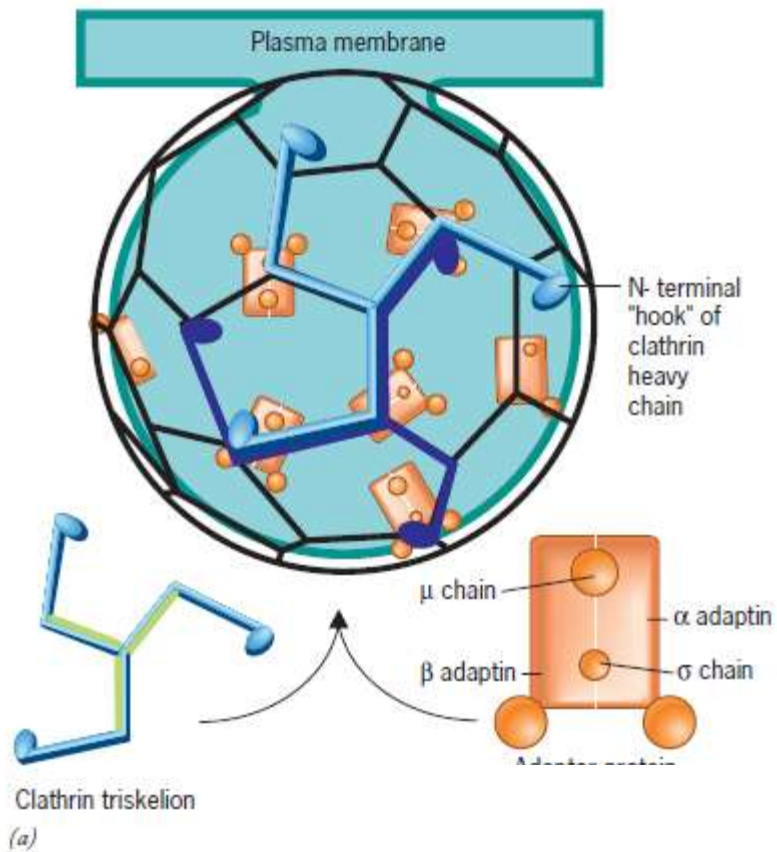


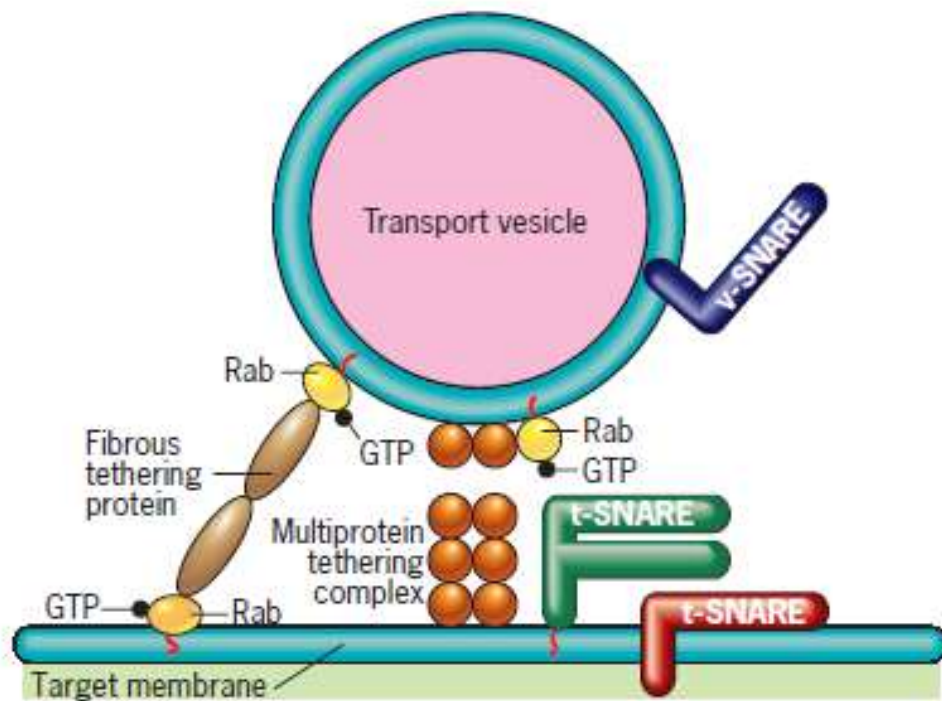
(c)



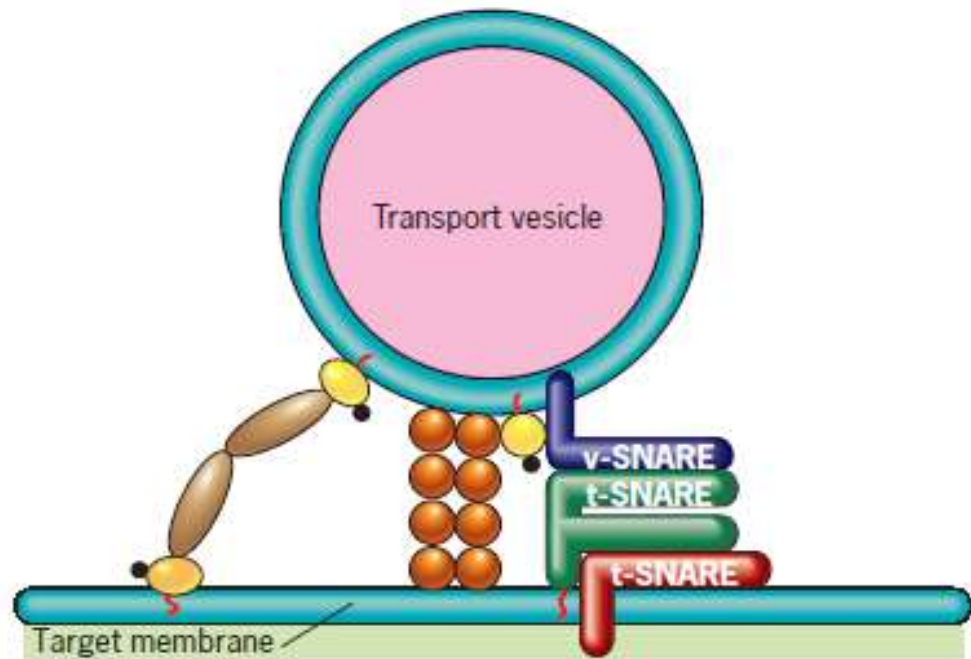
Exoplasmic face







(a) Tethering



(b) Docking

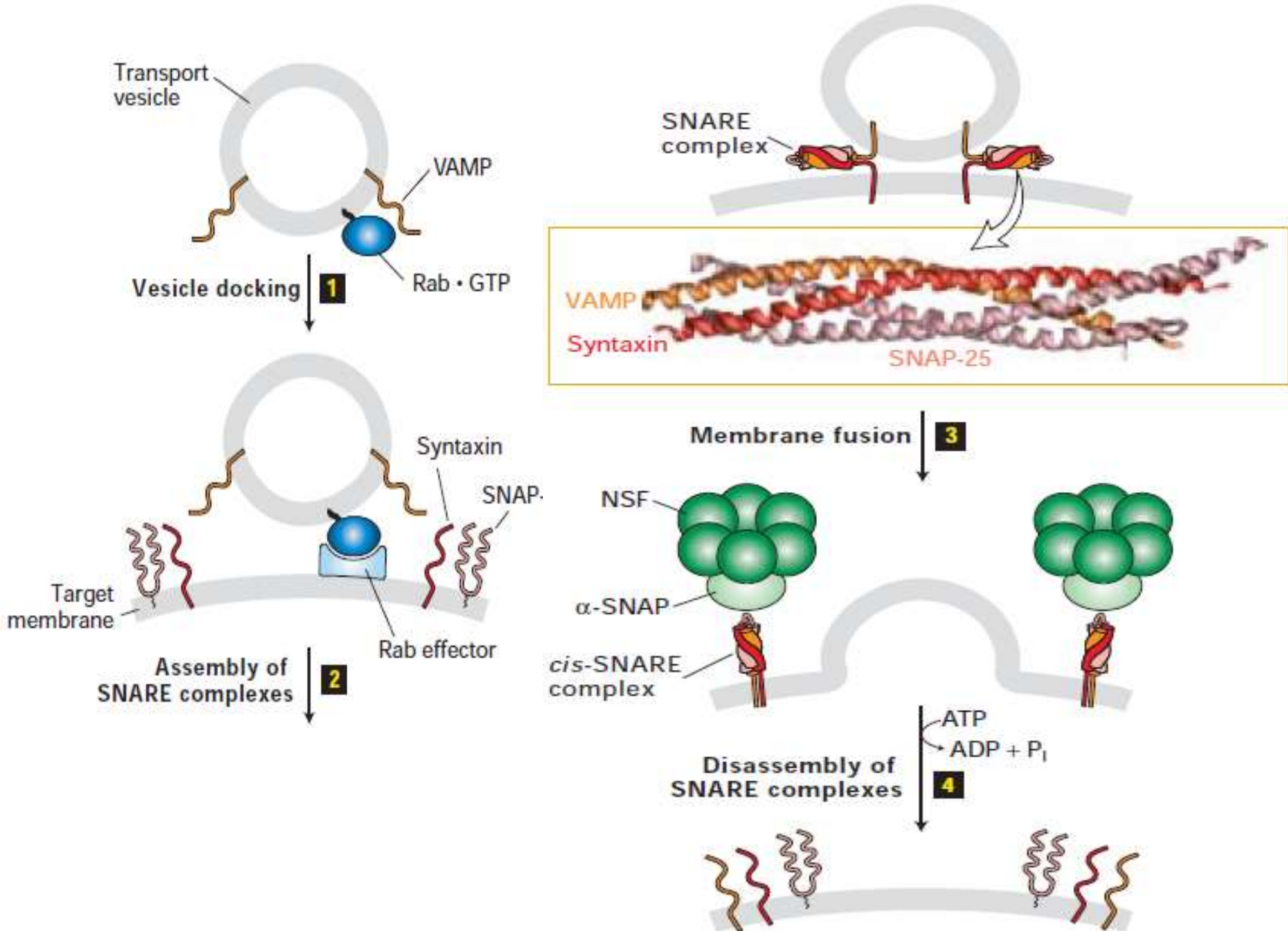
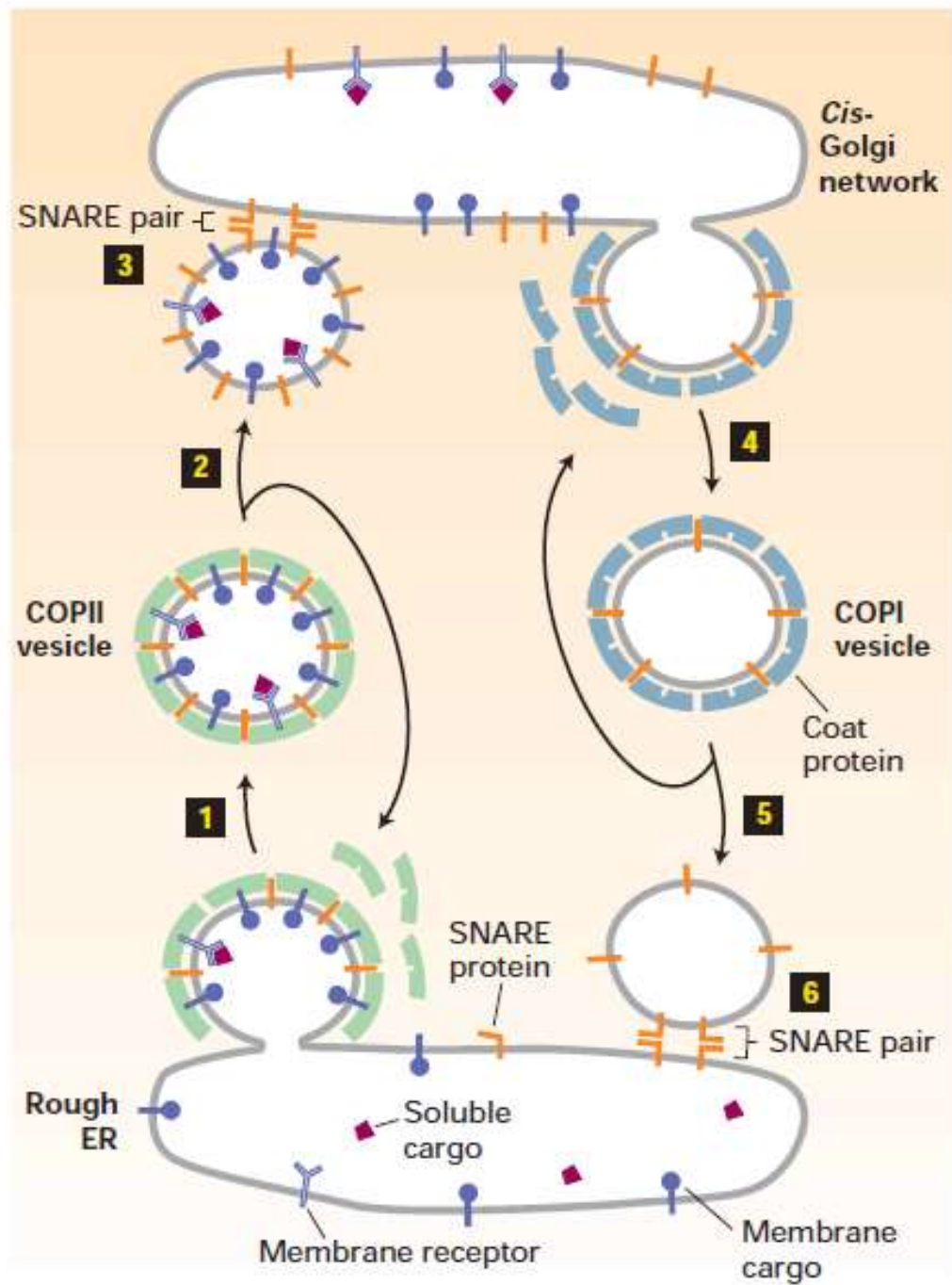
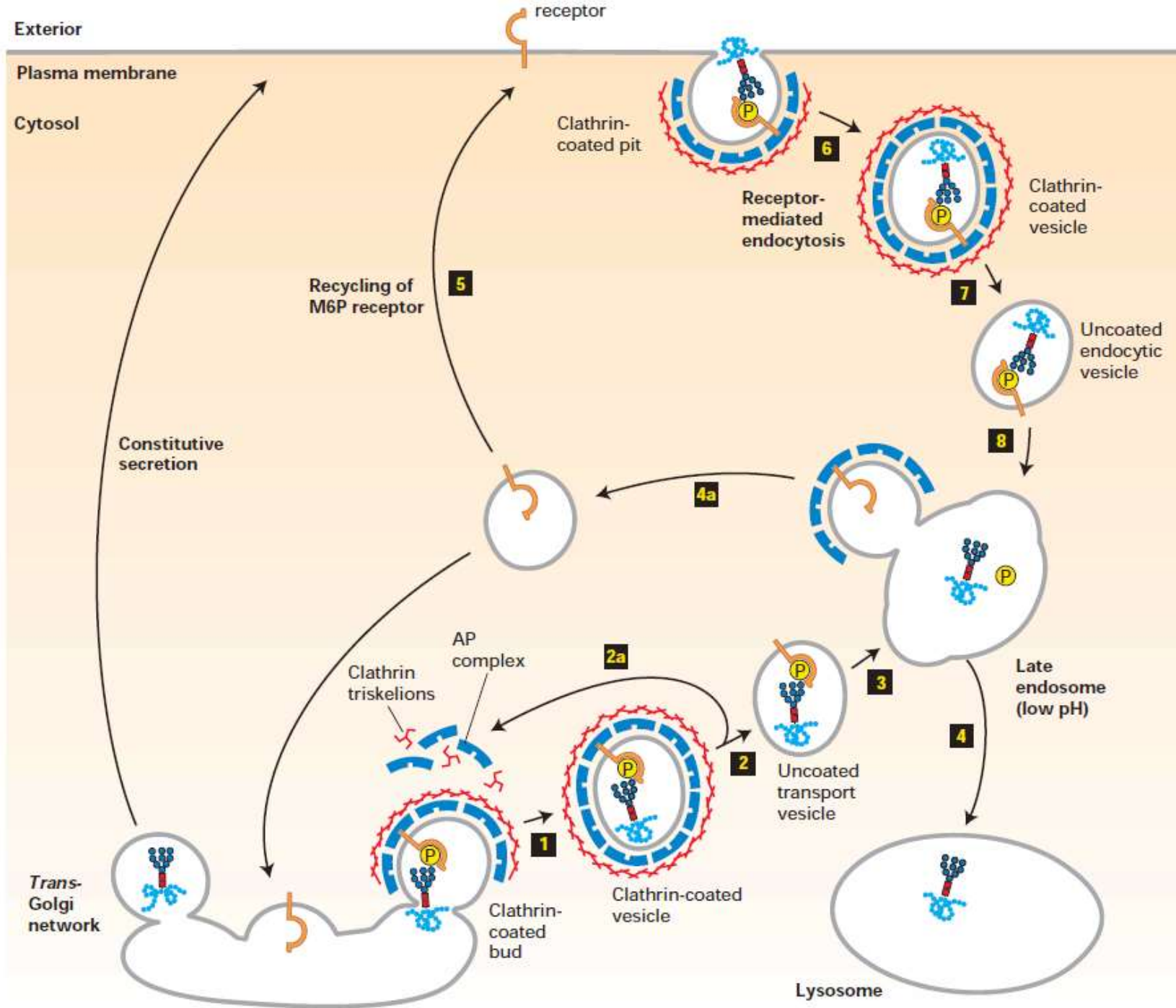


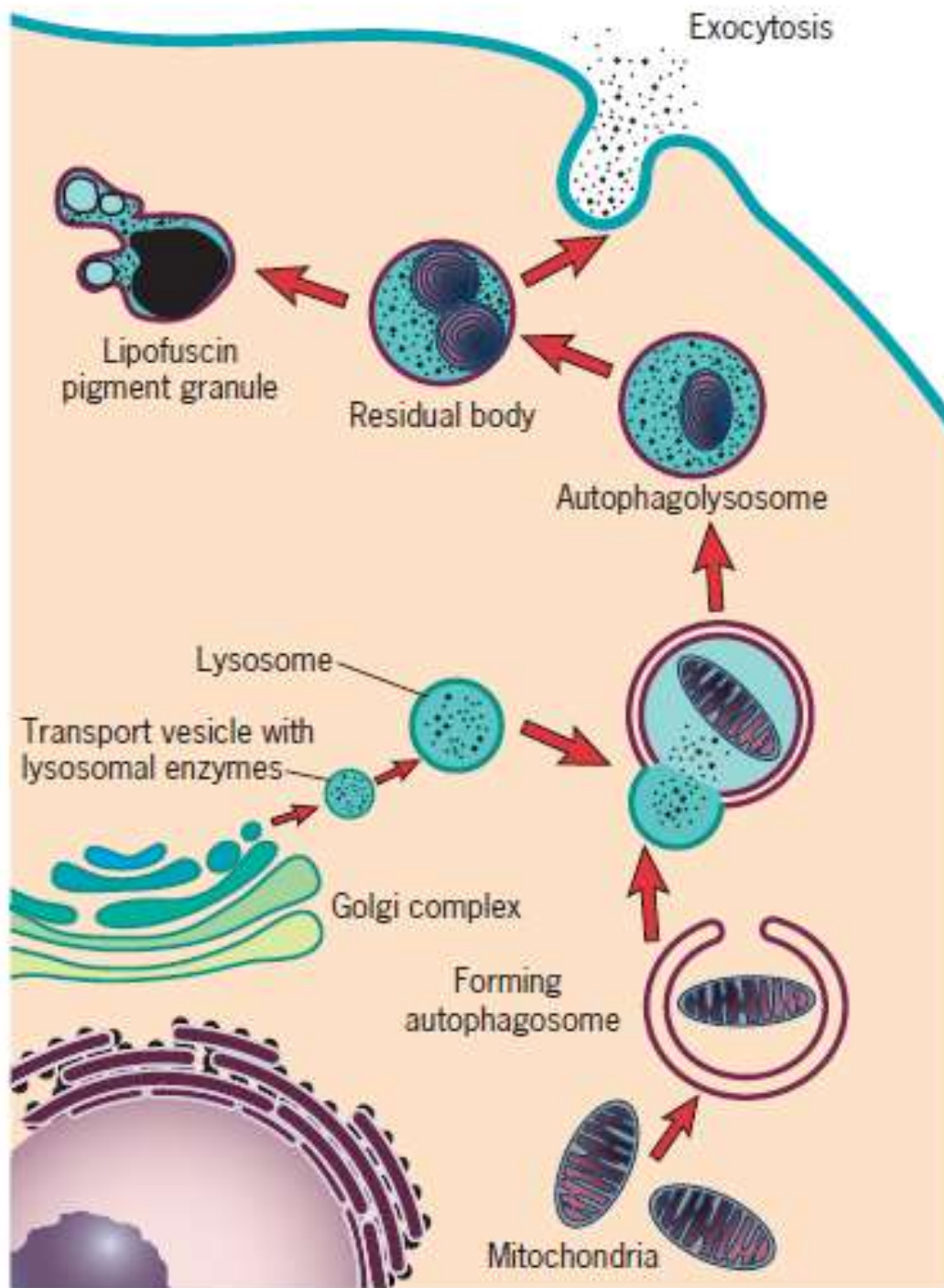
TABLE 17-2 Known Sorting Signals That Direct Proteins to Specific Transport Vesicles

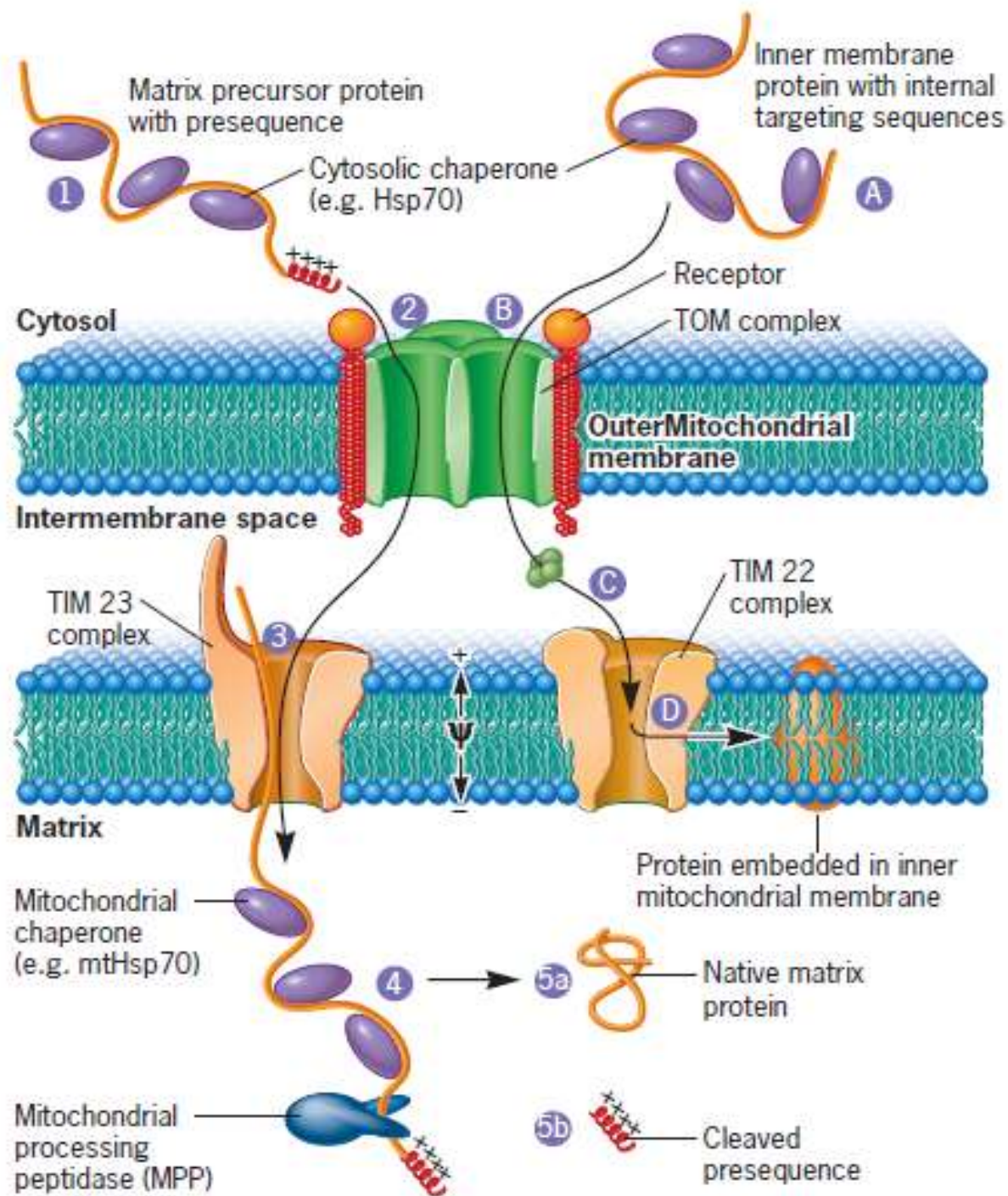
Signal Sequence*	Proteins with Signal	Signal Receptor	Vesicles That Incorporate Signal-bearing Protein
Lys-Asp-Glu-Leu (KDEL)	ER-resident luminal proteins	KDEL receptor in <i>cis</i> -Golgi membrane	COPI
Lys-Lys-X-X (KKXX)	ER-resident membrane proteins (cytosolic domain)	COPI α and β subunits	COPI
Di-acidic (e.g., Asp-X-Glu)	Cargo membrane proteins in ER (cytosolic domain)	COPII Sec24 subunit	COPII
Mannose 6-phosphate (M6P)	Soluble lysosomal enzymes after processing in <i>cis</i> -Golgi	M6P receptor in <i>trans</i> -Golgi membrane	Clathrin/AP1
	Secreted lysosomal enzymes	M6P receptor in plasma membrane	Clathrin/AP2
Asn-Pro-X-Tyr (NPXY)	LDL receptor in the plasma membrane (cytosolic domain)	AP2 complex	Clathrin/AP2
Tyr-X-X- Φ (YXX Φ)	Membrane proteins in <i>trans</i> -Golgi (cytosolic domain)	AP1 (μ 1 subunit)	Clathrin/AP1
	Plasma membrane proteins (cytosolic domain)	AP2 (μ 2 subunit)	Clathrin/AP2
Leu-Leu (LL)	Plasma membrane proteins (cytosolic domain)	AP2 complexes	Clathrin/AP2

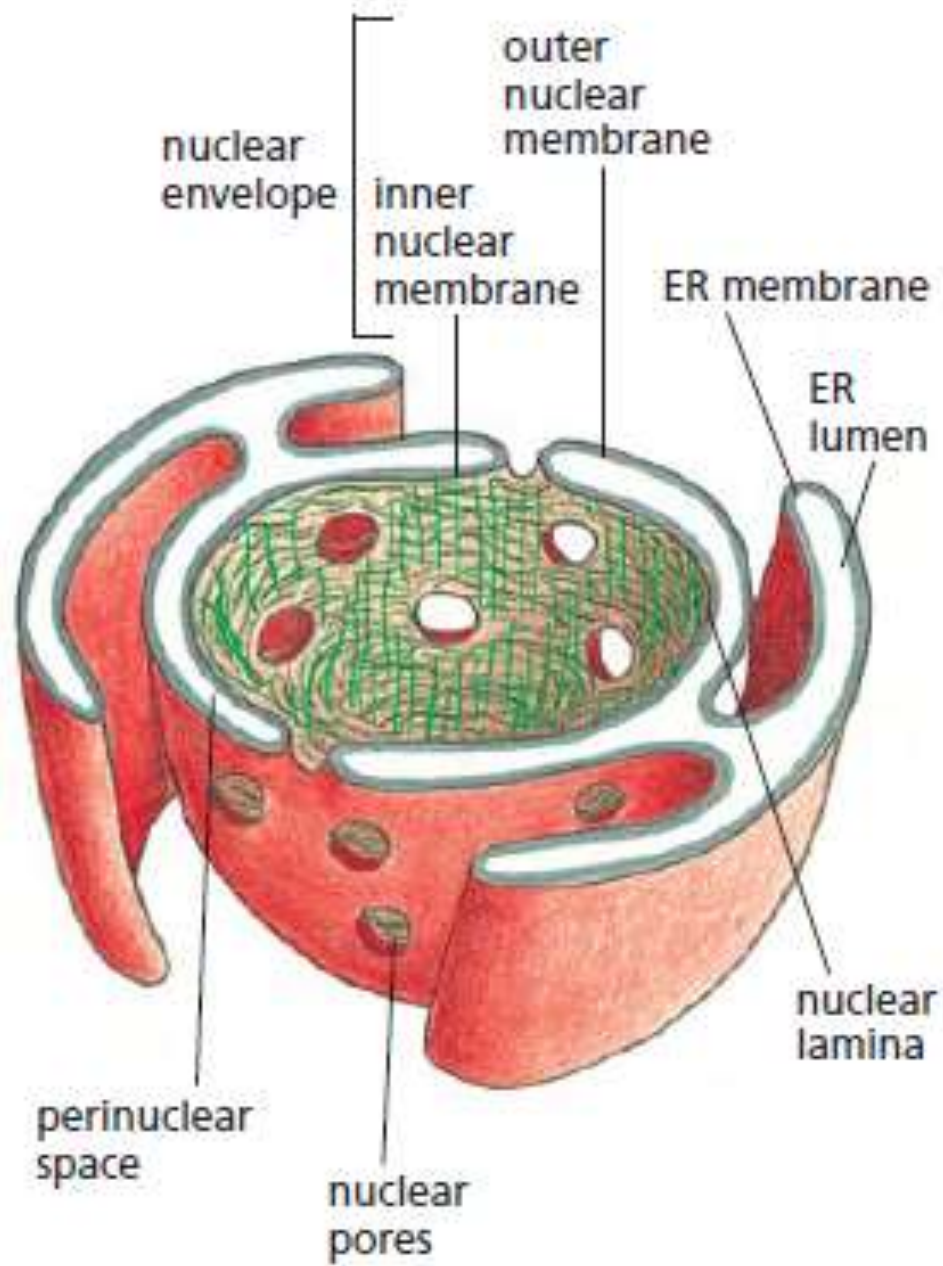
*X = any amino acid; Φ = hydrophobic amino acid. Single-letter amino acid abbreviations are in parentheses.

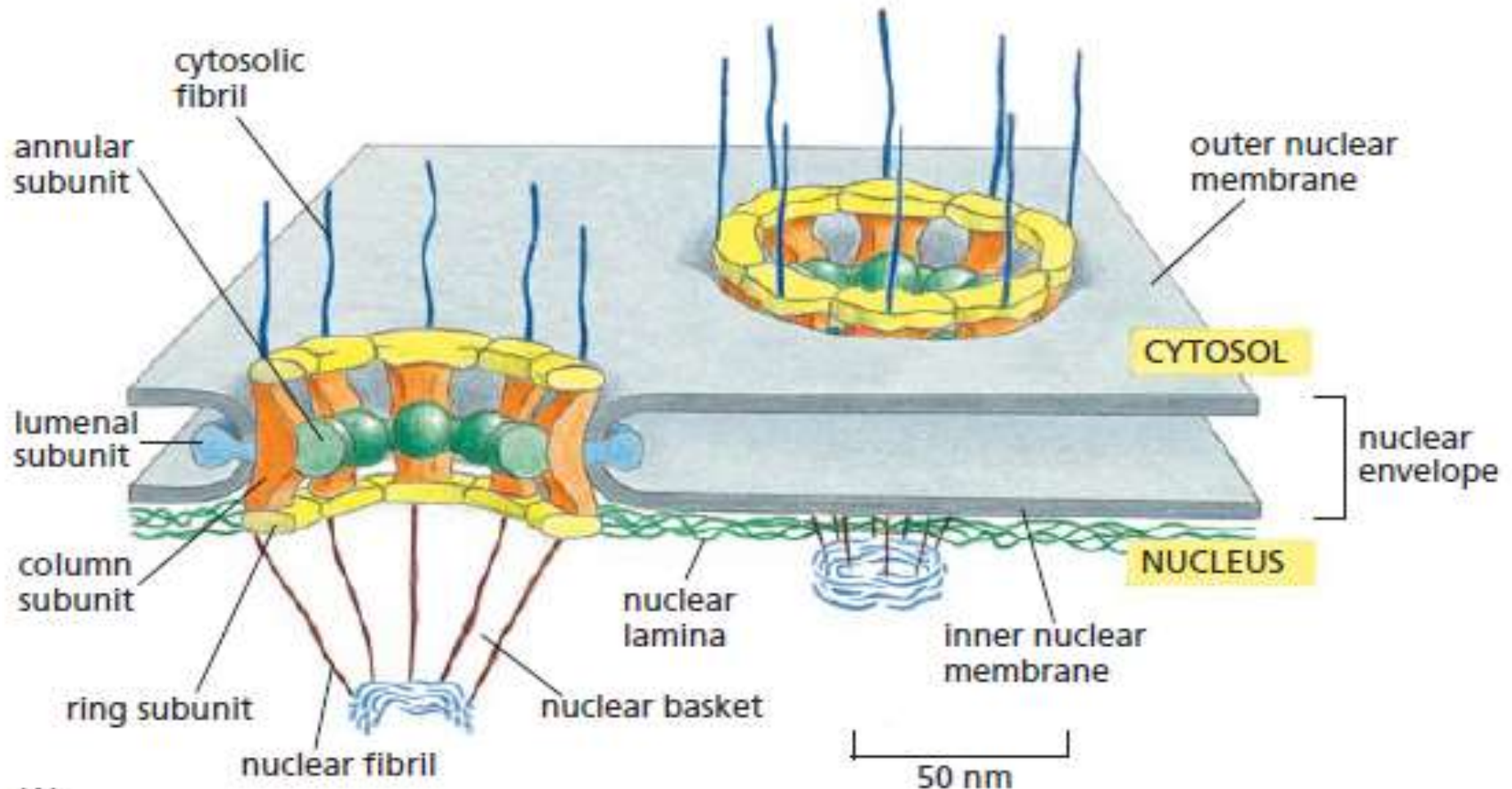




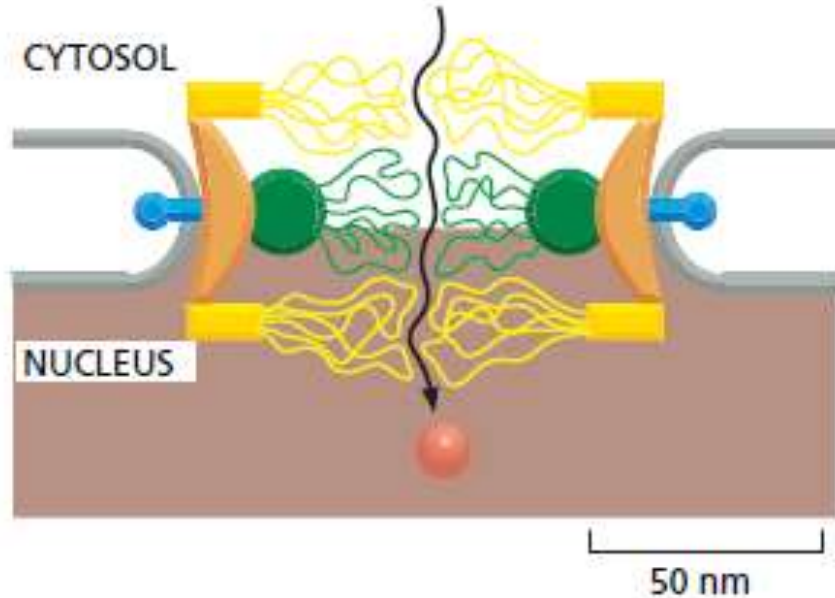








(A)



size of molecules
that enter nucleus
by free diffusion



size of macromolecules
that enter nucleus
by active transport

(A) LOCALIZATION OF T-ANTIGEN CONTAINING
ITS NORMAL NUCLEAR IMPORT SIGNAL

Pro — Pro — Lys — Lys — Lys — Arg — Lys — Val —



(B) LOCALIZATION OF T-ANTIGEN CONTAINING
A MUTATED NUCLEAR IMPORT SIGNAL

Pro — Pro — Lys — Thr — Lys — Arg — Lys — Val —



